Volume 9, Number 7, 2007 © Mary Ann Liebert, Inc. DOI: 10.1089/ars.2007.1629

Forum Review

Oxidative Stress, Glucose Metabolism, and the Prevention of Type 2 Diabetes: Pathophysiological Insights

SYED SHAH, MUHAMMAD IQBAL, JOCELYNE KARAM, MORO SALIFU, and SAMY I. McFARLANE

ABSTRACT

With the rising epidemic of type 2 diabetes worldwide, including the United States, the death and disability due to the suboptimal control of cardiovascular disease associated with this epidemic has made prevention of type 2 diabetes emerge as a primary strategic intervention. Several modalities have been assessed in large randomized controlled trials for diabetes prevention such as lifestyle interventions and various pharmacologic agents. Included in these agents are metformin, thiazolidinediones, acarbose, angiotensin converting enzyme inhibitors, as well as angiotensin receptor blockers. Abrogation of oxidative stress appears to be a common soil hypothesis that explains the favorable effects of these agents on glucose metabolism, including the prevention of diabetes and its complications. This comprehensive review highlights the role of oxidative stress in the pathogenesis of diabetes, with emphasis on the major clinical trials conducted on prevention of type 2 diabetes. *Antioxid. Redox Signal.* 9, 911–929.

INTRODUCTION

PIABETES MELLITUS is a major cardiovascular and renal risk factor and is ranked as the fifth leading cause of death worldwide (237). The increasing prevalence of diabetes has led to the increased risk of premature cardiovascular disease and death (31), prompting elaborate strategies to reduce its incidence. Despite public health efforts, diabetes remains a serious cause of morbidity and mortality. In 2002, diabetes was the sixth leading cause of death listed on United States death certificates and a leading cause of blindness and end stage renal disease in adults (42).

Estimates by the American Diabetes Association (ADA) indicate that 21 million people have diabetes and another 41 million are prediabetic, putting them at high risk for developing the disease in the near future. In fact, an estimated 1.5 million Americans develop diabetes every year. ADA also estimates the cost of diabetes, including such expenses as disability payments and lost days at work, to be at least \$132 billion a year, as of 2002 (42). The enormity of this epidemic with its

associated complications, morbidity, and mortality has initiated evidence-based efforts for its reduction (58, 63, 198).

The World Health Organization (WHO) estimates that >180 million people worldwide have diabetes, with its rate set to increase as childhood obesity rates soar worldwide. WHO estimates that 10% of school age children are overweight and >22 million children under the age of 5 years are obese or overweight. This number is likely to more than double by 2030. WHO also projects that death from diabetes will increase by >50% in the next 10 years without effective preventive strategies (118, 232).

Accumulating evidence indicates that oxidative stress plays a key role in the pathogenesis of diabetic microvascular and macrovascular complications (85), and endothelial dysfunction is an early marker of such damage (36, 85). Several *in vitro* (60, 271) and *in vivo* (19, 44, 178, 278) studies have shown that the acute effects of hyperglycemia can be reversed by antioxidants, thereby suggesting a role of free radicals in producing endothelial dysfunction caused by hyperglycemia (271).

Division of Endocrinology, Diabetes and Hypertension, Department of Internal Medicine, SUNY Downstate and Kings County Hospital Center, Brooklyn, New York.

This is a comprehensive review of oxidative stress and its role in diabetes pathogenesis, highlighting the role of oxidative stress in light of several clinical trials conducted on prevention of type 2 diabetes.

OXIDATIVE STRESS

Oxidative stress is defined as tissue injury resulting from a disturbance in the equilibrium between the production of reactive oxygen species (ROS), also known as free radicals, and antioxidant defense mechanisms (24, 103, 256). Under physiologic conditions, the antioxidant defenses are able to protect against the deleterious effects of free radicals, but under conditions where either the free radical formation is increased or the antioxidant defenses are inactivated, accumulation of free radicals ensues, leading to cellular and tissue damages (24, 96, 103, 256).

Although the history of oxygen toxicity in laboratory animals dates back to 1878 (148), the first description of free radicals came in 1894, when presumably, Fenton reported the first free radical reaction (77). Oblivious to the existence of a moiety called "free radicals" at the time, Fenton described the generation of a hydroxyl free radical as a result of the reduction of hydrogen peroxide with ferrous iron in a solution of tartaric acid—a classic mechanism known as the "Fenton reaction".

$$Fe^{2+} + H_2O_2 \rightarrow Fe^{3+} + HO^- + {}^{\bullet}OH$$

The first organic free radical, however, was described by Moses Gomberg, in 1900, when he published the results on the reaction of triphenylmethyl halides with metals leading to the formation of triphenylmethyl radical (Ph., C.) (87).

The role of free radicals in disease processes such as cell injury, cancer, and aging, was originally recognized by Harman (106, 107) who hypothesized the *in vivo* generation of free radicals as a major contributor in the pathogenesis of disease. The discovery of superoxide dismutase, in 1969, established the role of free radicals in biological systems (148). Subsequently, extensive research has established the importance of free radicals in aging (106, 107), as well as several pathological conditions including coronary artery disease (CAD), stroke, ischemic dementia (86, 217, 262), carcinogenesis (8, 45), neurodegenerative disorders (149), pulmonary disease (226), renal disease (181) and diabetes mellitus, which will be discussed in detail.

Free radicals (reactive oxygen species: ROS)

A free radical is defined as any atom or molecule that contains one or more unpaired electrons (101). The presence of unpaired electrons increases the reactivity of an atom or molecule, thereby making it much more reactive than a corresponding nonradical. Free radicals have been known to have deleterious as well as beneficial effects (286).

Free radicals are generated in huge amounts as byproducts of common physiologic reactions, as well as end products for specific defense purposes, such as neutrophil activation. Additional sources of free radical acquisition include ozone, nitrogen dioxide, and electromagnetic radiation (24).

Numerous free radicals are known to perform various functions in the body (Fig. 1). Common examples of free radicals include but are not limited to hydroxyl (*OH), superoxide (O₂-*), nitric oxide (NO*), hydrogen peroxide (H₂O₂), peroxyl (ROO*), alkoxyl (RO*), thiyl (RS*), and peroxynitrite (ONOO-) (24, 148, 292). Superoxide, nitric oxide, and hydrogen peroxide play an important role in normal physiology, but at the same time are well known to be responsible for accelerating the aging process and cell degeneration in disease states (292).

The earliest description of hydroxyl ('OH) radical generation comes from the Haber–Weiss reaction (102), described in the 1930s, which showed the conversion of super-oxide to the hydroxyl radical (Fig. 1). This extremely potent radical has a half-life, in aqueous solution, of less than 1 ns (218). It is generated in the human body by splitting of water molecules as a result of ionizing radiations from the environment and photolytic decomposition of alkylhydroperoxides (101, 286).

Superoxide (O_2^-) is generated in the body by mitochondria when reduced nicotinamide adenine dinucleotide (NADH) is oxidized to nicotinamide adenine dinucleotide (NAD)⁺. Superoxide (O_2^-) could also be a byproduct of direct reactions of various molecules with oxygen, such as catecholamines and tetrahydrofolates (103, 166, 292) (Fig. 1). Although generally regarded as weakly reactive, superoxide (O_2^-) plays an important role in the immune system, as it is produced by activated phagocytes.

Nitric oxide (NO*) is generated through nitric oxide synthase (NOS) and is present in the body in large quantities, acting as a major signaling molecule at the endothelial level in multiple physiological processes (7, 23, 59). NO* is also poorly reactive and has a half-life of only a few seconds in an aqueous environment (50).

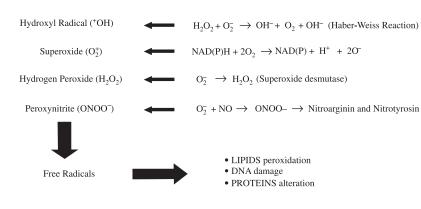


FIG. 1. Origins of main oxygen and organic free radicals and effects on cells.

There is strong evidence supporting the role of metals such as iron, copper, chromium, arsenic, nickel, cadmium, cobalt, and vanadium, causing free radical-induced damage in biological systems, thereby enhancing the process of carcinogenicity (163, 286).

Free radical-induced cell injury

The presence of an odd number of electrons makes free radicals unstable, thereby rendering them highly reactive. Although this reactivity varies between different radicals, free radicals continuously react to the nearby molecules, accepting or donating electrons, in order to achieve a stable state. The majority of these reactions are either between a radical and a nonradical, and a few that occur between two free radicals.

Common examples of reactions between radicals and non-radicals include lipid peroxidation, protein damage (24, 100, 261), and DNA damage that can result from the presence of free radicals in the immediate vicinity of DNA, for example, the conversion of guanine into 8-hyroxyguanine and other products, by hydroxyl radicals (21, 29, 61, 100). The combination of superoxide (O_2^-) and nitric oxide (NO*), leading to the formation of peroxynitrite (ONOO*-), however, is an example of a reaction between two radicals (Fig. 1). Peroxynitrite (ONOO*-), at physiological pH, is injurious to proteins directly and it further decomposes into toxic products like nitrogen dioxide gas (NO2*), hydroxyl radical (*OH), and nitronium ion (NO2*) (20).

Oxidative stress is a state of imbalance between the generation of free radicals and the antioxidant defense mechanisms. Antioxidants enhance the indigenous ability to protect against the free radical damage. These antioxidants are defined as any compound that can donate at least one hydrogen atom to a free radical, resulting in the termination of radical chain reactions (292).

Examples of antioxidants include Vitamin E, beta-carotene, co-enzyme Q, and the enzymes dismutase, peroxidase, and catalase (24). The antioxidants, such as ceruloplasmin, transferrin, and albumin, defend against the harmful effects of free radicals by preventing the initiation of the free radical chain reaction due to their ability to bind metal ions (263, 292).

Measurement of oxidative stress

As discussed earlier, there is strong evidence that oxidative stress plays a role in the pathogenesis of different diseases (8, 45, 86, 106, 107, 149, 181, 217, 226, 262), including diabetes and its complications, but to date there is no direct measure of oxidative stress in biological systems (130). There are several biomarkers that have been identified as measure of oxidative damage at the molecular level, such as isoprostane assays (40, 195), transcriptional activation assays (221), glutathione assays (247), malondialdehyde (MDA), thiobarbituric acid-reactive substances (TBARS) and lipid peroxidation (126), superoxide dismutase, 8-hydroxy-deoxyguanosine (280), catalase (306), 5,5-dimethyl-1-pyrroline-N-oxide (94), human myeloperoxidase (144), human plasma lactoferrin (13), among several others. New and improved methods, such as electron spin resonance (ESR) (205), are being developed and perfected to measure oxidative stress more reliably and accurately. Recently, researchers have shown increased levels of xanthine and NADPH oxidase in human coronary artery disease using ESR (260).

A comprehensive discussion on the measurement of oxidative stress is beyond the scope of this review.

Human disease and oxidative stress

Free radical and oxidative stress have become an integral part of understanding the underlying mechanism of disease in today's modern medicine. With the advances in cellular and molecular research, there is considerable and increasing evidence linking oxidative stress and various human diseases. This is hardly surprising given the fact that oxidative metabolism is an indispensable part of cell physiology. In fact, oxidative stress is a well-documented component of several diseases such as acquired immunodeficiency syndrome (AIDS) (78), adult respiratory distress syndrome (ARDS) (89), Alzheimer's disease (169), amyotrophic lateral sclerosis (52), arthritis (285), diabetes mellitus (186), emphysema (294), gastric ulcers (55), glomerulonephritis (254), heart disease (212), hemochromatosis (116), hypertension (141), intestinal ischemia (216), lupus erythematosus (192), multiple sclerosis (279), muscular dystrophy (227), organ transplantation (25, 199), Parkinson disease (52), preeclampsia (120), stroke (15), vasculitis (295), and many others.

OXIDATIVE STRESS AND GLUCOSE METABOLISM

Role of oxidative stress in the etiology of diabetes

Diabetes mellitus is a metabolic disorder characterized by hyperglycemia resulting from absolute or relative deficiency and/or insensitivity to endogenous insulin. Traditionally, it has been classified into two forms, type 1, which is caused by autoimmune destruction of pancreatic β -cells, and type 2, which is known to be multifactorial, resulting from combination of various factors such as impaired fatty acid metabolism, central fat deposition leading to insulin resistance (186), β -cell secretory defect, and obesity (17, 80). This view, however, is currently being challenged by the accelerator hypothesis that argues that type-1 and type-2 diabetes are the same disorder of insulin resistance set against different genetic backgrounds (147).

Evidence has long existed regarding the relationship between oxidative stress and diabetes mellitus (207, 248, 302). The role of oxidative stress in the etiology of diabetes was recognized in the early 1980s, when streptozocin and alloxan (175, 187) were used in experimental animals to induce diabetes. These agents were shown to result in diabetes via mechanisms involving either an increase in the production of free radicals or a decrease in antioxidant defenses (302).

Simoneau and colleagues, in 1995, reported for the first time the relationship between mitochondrial dysfunction and diabetes (258). Since then, research has suggested the role of oxidative stress and mitochondrial dysfunction in the pathogenesis of type 2 diabetes, specifically in intrauterine growth retardation (IUGR) (139, 257). In growth-retarded fetuses, low oxygen levels generate the production of free radicals due to decreased activity of the electron transport chains (46, 70), leading to DNA,

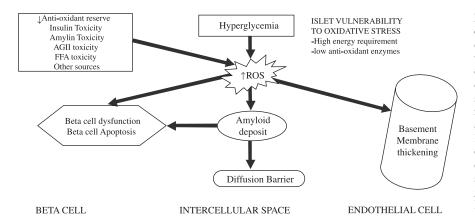


FIG. 2. Effect of oxidative stress on islet beta cells. The islet beta cells are particularly vulnerable to the effect of reactive oxygen species originating from hyperglycemia and other related factors. Oxidative stress may cause an unfolding of the native secondary structure of amylin with subsequent refolding leading to deposit of islet amyloid, creating a secretory and absorptive defect within the islet in addition to space occupying lesion. AGII, angiotensin II; FFA, free fatty acids; ROS, reactive oxygen species.

protein, and mitochondrial damage—decreasing energy production (257). This mitochondrial dysfunction has lethal effects on β -cell function, which has a high energy requirement and depends upon normal ATP production for insulin proliferation and secretion (10, 176, 188, 200, 204, 215, 252). Another reason for the increased β -cell vulnerability comes from the fact that antioxidant enzyme expression is very low in pancreatic islets (162, 275, 276). Furthermore, there is considerable evidence for free radical-induced inhibition of glucose-mediated insulin secretion, apoptosis, and diminished β -cell gene expression (62, 129, 135, 136, 137, 171, 194, 244) (Fig. 2).

Evidence suggests that mitochondrial dysfunction can further lead to oxidative stress via various mechanisms (257). Animal studies have shown that muscle mitochondria in IUGR have chronically decreased ATP production, causing compromised glucose transporter isoform-4 (GLUT-4)

recruitment to the cell surface, glucose transport, and glycogen synthesis. This further leads to insulin resistance and hyperglycemia of type 2 diabetes (253). Human studies using $_{13}$ C and $_{31}$ P magnetic resonance spectroscopy in offsprings of parents with type 2 diabetes have suggested mitochondrial dysfunction as a possible contributing factor to the development of diabetes (222).

The polyol pathway is an established contributor to oxidative stress (159). The enzyme aldose reductase, which is responsible for detoxification of aldehydes to alcohols, in the presence of cellular hyperglycemia, reduces glucose to sorbitol, which is oxidized to fructose, consuming large quantities of NADPH, leading to its depletion (Fig. 3). The depletion of NADPH leads to oxidative stress due to the fact that it is a major cofactor for glutathione regeneration and a known antioxidant defense at the cellular level. Additionally,

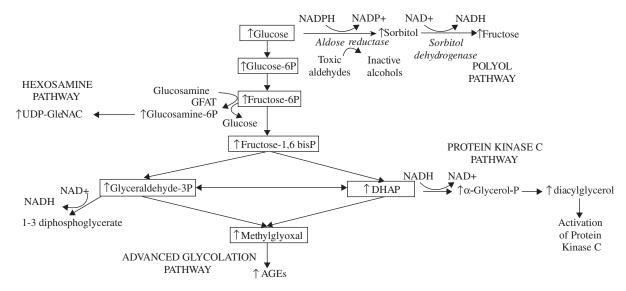


FIG. 3. Major pathways of hyperglycemia-induced oxidative stress. Hyperglycemia leads to increased flux through polyol pathway with subsequent NADPH depletion; increased flux through hexosamine pathway with subsequent gene expression alterations, increased intracellular production of AGE precursors with subsequent direct intra- and extracellular pathophysiologic consequences, and activated protein kinase C affecting genes expression. AGE, advanced glycation end products; DHAP, dihydroxyacetone phosphate; GFAT, glutamine:fructose-6 phosphate amidotransferase; NAD, nicotinamide-adenine dinucleotide; NADP+, oxidized nicotinamide-adenine dinucleotide phosphate; NADPH, nicotinamide-adenine dinucleotide phosphate; UDP Glc-NAC, uridine diphosphate *N*-acetyl glucosamine.

FIG. 4. The glutathione oxidation reduction cycle. H₂O₂, hydrogen peroxide; NADP+, oxidized nicotinamide-adenine dinucleotide phosphate; NADPH, nicotinamide-adenine dinucleotide phosphate; O₂*, superoxide.

measures that inhibit the polyol pathway delay the evolution of diabetic complications (67) (Fig. 4).

dimutase

In addition to the evidence discussed above, implicating oxidative stress as an underlying factor in the pathogenesis of diabetes, there is considerable evidence regarding indirect role of oxidative stress in the etiology of this disease. Chronic hyperglycemia, besides increasing free radical production, is associated with increased production of advanced glycation end products (AGE) and lipid peroxidation products, as well as decreased antioxidant defenses (71, 72) (Fig. 3). Furthermore, animal studies have elucidated a protective role of antioxidants against free radical-mediated injury during short-term hyperglycemic states (97). Numerous nutritional supplements and pharmacologic agents have been shown to improve both insulin resistance and oxidative stress in hyperglycemic states (255).

In diabetes, there is increased generation of pro-oxidants such as cytokines and peroxides (172, 270). Pancreatic islets, having intrinsically low levels of antioxidant enzyme expression (90, 275, 276), are especially susceptible to free radical damage. This, coupled with adverse effects of lipid peroxidation, results in increased islet apoptosis (236) (Fig. 2).

These data collectively highlight the link between diabetes and the imbalance of free radical production and antioxidant defense mechanism. It also provides evidence that oxidative stress plays a key role in diabetic complications (18).

Oxidative stress and metabolic syndrome

Although metabolic syndrome is not included in the defining criteria for diabetes, most people with diabetes have metabolic syndrome and most patients with metabolic syndrome are at risk for developing diabetes.

Metabolic syndrome, initially defined by Reaven in 1988 (229), consists of obesity (abdominal circumference men >40 in and women >35 in), impaired glucose tolerance (fasting plasma glucose ≥100 mg/dL), hypertension (blood pressure ≥130/85 mm Hg), and dyslipidemia characterized by elevated triglycerides (≥150 mg/dL) and low high density lipoprotein (<40 mg/dL) levels (93, 229). All of the individual features of metabolic syndrome are considered to be significant risk factors for cardiovascular disease. This association has recently

been termed as cardiometabolic syndrome (CMS), which includes congestive heart failure (CHF), coronary heart disease (CHD), and stroke (108, 110, 158, 202). One of the mechanisms by which CMS results in myocardial injury involves the production of reactive oxygen species and decreased NO resulting from endothelial NO synthase (eNOS) uncoupling (108, 109).

Obesity, one of the key factors of metabolic syndrome, leads to increased oxidative stress caused by reduced availability of NO, thereby leading to increased vascular tone and subsequent hypertension (54). Furthermore, obesity also leads to increased asymmetric dimethylarginine (ADMA) concentrations, which results in eNOS dysfunction as a consequence of its competition for the substrate L-arginine. This causes eNOS uncoupling with increased superoxide production, decreased endothelial nitric oxide, and endothelial dysfunction (111, 165).

Recently it has been shown that there is a positive correlation between body mass index and systemic oxidative stress (140, 211). Furukawa *et al.* demonstrated that a major source of free radicals in plasma is adipose tissue, which subsequently leads to insulin resistance in skeletal muscle and adipose tissue and impaired insulin secretion by β -cells (82). The relationship between obesity and oxidative stress also holds true in nondiabetic subjects (82), making obesity a major player in the pathologic outcomes of metabolic syndrome.

Evidence for the pathogenic role of oxidative stress in essential hypertension is inconclusive (154, 157, 174, 197, 305) despite considerable data supporting the presence of increased free radical production in hypertension. Elevated levels of superoxide, hydrogen peroxide, lipid peroxides, plasma hydrogen peroxide, and decreased superoxide dismutase (SOD) have been observed in hypertensive patients compared to normotensive patients (28, 154, 157, 219).

Free radicals have reactivity with almost all biological substances, the most susceptible of which are polyunsaturated fatty acids which are constituents of cell membranes. These react with the free radicals, leading to lipid peroxidation, a biomarker of oxidative stress (126, 207, 302). Lipid peroxidation is the first step in the generation of oxidized LDL and results in increased production of free radicals and causes platelet activation and increased cardiovascular risk. Oxidized LDL also causes

endothelial damage, macrophage activation and impaired vasodilation (84, 161, 284). Furthermore, in metabolic syndrome, there is impaired antioxidative and anti-inflammatory activity of HDL molecules, leading to elevated systemic oxidative stress as measured by plasma 8-isoprostane levels (105).

The notion that oxidative stress is a major contributor in metabolic syndrome is further supported by the fact that four out of the five criteria (as defined by NCEP/ATP III) are independently associated with increased levels of oxidative stress (14, 140, 207, 230).

Role of oxidative stress in diabetic complications

Hyperglycemia, a characteristic feature of diabetes, predisposes to vascular complications, both microvascular as well as macrovascular, and an early indicator of such damage is endothelial dysfunction (36, 85). These complications result from diverse mechanisms, and oxidative stress has now been suggested to be a common pathway linking these mechanisms to the pathogenesis of diabetic complications (91, 203, 239).

There are four major pathways linking oxidative stress as a contributing factor to the complications of hyperglycemia. These include increased polyol pathway flux, increased advanced glycosylation end (AGE) product formation, activation of protein kinase C, and increased hexosamine pathway flux (33, 64) (Fig. 3). These processes also cause endothelial dysfunction that enhances the development and progression of diabetic complications (60). Furthermore, there is a well-established relationship between diabetes and vascular disease (138). Several clinical trials have established long-term glycemic control as an independent predictor of diabetic vascular complications (156, 272, 283). In addition, insulin has been shown to have anti-inflammatory and antioxidant effects, as evident by its ability to suppress free radical production and p47phox expression—a component of the enzyme complex

that produces superoxide free radical (53). These beneficial cardiovascular effects are now being studied in large trials to assess if early insulin therapy would decrease cardiovascular disease in diabetes. Among these is the Outcomes Reduction with Initial Glargine Intervention (ORIGIN) (201).

Cardiovascular disease

Cardiovascular disease is the major cause of morbidity and mortality in diabetes (43). Furthermore, diabetes is now considered a cardiovascular risk equivalent (1, 112, 190). Therefore, diabetic patients who develop myocardial infarctions have poorer prognosis than nondiabetic individuals (79, 98, 119, 125). Diabetes also increases the risk of stroke in diabetic patients by two- to four-fold compared to nondiabetic individuals (99, 177).

The common link in these complications is accelerated atherosclerosis in diabetes (12, 128) where the primary triggers of atherogenesis are insulin resistance and hyperlipidemia, both of which are features of diabetes (12). One of the key mechanisms of premature atherosclerosis in diabetes is the oxidation of low density lipoprotein (LDL) leading to oxidative stress (228) (Fig. 5). Multiple studies have documented the link between diabetes and enhanced LDL oxidative vulnerability (164, 170).

There is increased vascular endothelial free radical production in diabetic subjects, which provides the milieu enhancing the oxidation of LDL (95). As elucidated to earlier, endothelial dysfunction is one of the cardinal features of premature diabetic complications (36, 85, 127). Furthermore, antioxidants have been shown to improve endothelial function, providing indirect evidence for a pathogenic role of oxidative stress in diabetic endothelial dysfunction (224, 259). Increased superoxide production, caused by NADPH oxidase and uncoupled eNOS, is a major contributor to this free radical-mediated injury (128) (Fig. 5).

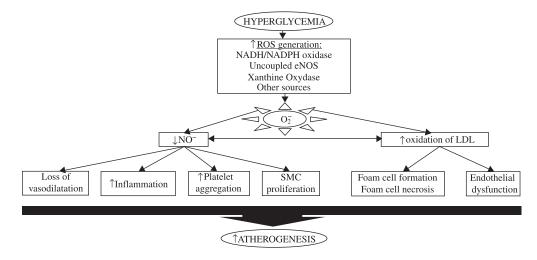


FIG. 5. Role of oxidative stress in diabetes predisposing to atherogenesis. Generation of excess of ROS in diabetes is multifactorial and favors superoxide formation in the superoxide–nitric oxide (NO) balance normally maintaining endothelial function. This oxidative stress will result in increased LDL oxidation and loss of NO beneficial effects resulting in atherosclerosis. eNOS, endothelial nitric oxide synthase; LDL, low-density lipoprotein; NO, nitric oxide; O₂, superoxide; ROS, reactive oxygen species; SMC, smooth muscle cell.

There is strong evidence linking the increased production of NADPH oxidase in presence of hyperglycemia, oxidized LDL, AGE, and free fatty acids (FFA) (122, 123, 206, 298). Moreover, NADPH oxidase, isolated from vessels of diabetic patients, is more active as evident by increased expression of its subunits, especially p22 phox and p47phox (95). As mentioned earlier, uncoupling of eNOS is an important contributor to the free radical production. Hyperglycemia, like obesity, also results in accumulation of ADMA, which leads to increased eNOS, thereby causing superoxide production (165).

The migration, activation, and release of cytokines by monocytes have a well-documented role in the formation and progression of atherosclerotic plaques. These activated monocytes further develop into macrophages to become foam cells, which are rich sources of inflammatory mediators and free radicals (240). The increased expression of monocyte chemoattractant protein (MCP-1) is known to increase the adhesion of monocytes in hyperglycemic states (269). Dhindsa *et al.* showed that monocyte-mediated free radical generation is increased in healthy volunteers after a single oral glucose dose (57). In addition, monocytes isolated from patients with poorly controlled diabetes, via various mechanisms, increase superoxide production (114, 289).

Dysfunctional vascular smooth muscle in diabetics enhances their susceptibility to premature atherosclerosis (177, 195). Vascular smooth muscle cells (VSMC) produce increased superoxide under hyperglycemic conditions (122, 123, 160). There is enhanced atherosclerotic lesion formation in diabetic patients as a result of the inhibition of vasodilation, caused by production of oxidative stress by VSMC, and increased migration of VSMC into atherosclerotic lesions (128, 267). Oxidative stress, via hydrogen peroxide and oxidized LDL, leads to VSMC necrosis and apoptosis in diabetic patients' coronary arteries and aortas (117, 220). The resultant VSMC death is now suggested to play a vital role in plaque instability and rupture (81).

A distinct but clinically relevant entity, diabetic cardiomyopathy, has been recognized based on clinical, epidemiologic, and pathological data (73, 241). Although the exact cause of diabetic cardiomyopathy remains unclear (27), recent studies suggest a pathogenic role of free radical-mediated apoptosis that leads to the cascade of events eventually causing cardiomyopathy (37, 38). Moreover, considerable similarities exist between cardiomyopathy induced by nitric oxide inhibition and cardiomyopathy induced by the combination of hypertension and diabetes, suggesting an important role of eNOS and eNO in the pathogenesis of diabetic cardiomyopathy (110, 245).

Nephropathy

Diabetic nephropathy is characterized by persistent albuminuria, confirmed on at least two occasions 3–6 months apart, declining glomerular filtration rate (GFR), and hypertension. Diabetic nephropathy develops in ~35% of patients with type 1 diabetes mellitus and 15%–20% of patients with type 2 diabetes mellitus, and is the leading cause of kidney failure, accounting for 44% of new cases in 2002 (41, 111).

Although review of the literature reveals many mechanisms that have been proposed to play a role in diabetic nephropathy, some important pathogenic mechanisms include, but are not limited to, the formation of free radicals, increased formation of AGE, the activation of protein kinase C (PKC), increased growth factor activity, the activation of cytokines, and decreased glycosaminoglycan content in basement membranes (39).

Oxidative stress injury in diabetic nephropathy is potentially mediated by multiple factors, especially free radical generation due to mitochondrial dysfunction and decreased activity of protective mechanisms (22, 257) (Fig. 6). Nitric oxide is excessively active in diabetics with microalbuminuria as compared to nonalbuminuric patients and was found to be related to GFR (49). Mouse studies of inducible nitric oxide synthase (NOS) suggest that NO modulates glomerulosclerosis and tubulointerstitial fibrosis (281).

Additional evidence comes from studies showing increased levels of 8-hydroxy-2'-deoxyguanosine (8-OHdG), indicating oxidative mitochondrial DNA damage and deletion, in rat kidneys (133). Similar studies in humans have

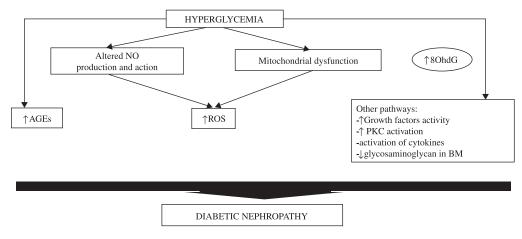


FIG. 6. Implications of oxidative stress in pathophysiology of diabetic nephropathy. AGEs, advanced glycation end products; BM, basement membrane; NO, nitric oxide; 8-OHdG, 8-hydroxy-2'-deoxyguanosine; PKC, protein kinase C; ROS, reactive oxygen species.

demonstrated increased urinary levels of 8-OHdG in type 2 diabetics as compared to controls, and this increase was directly proportional to the severity of glomerular and tubulointerstitial lesions (134). A 5-year prospective Japanese trial, concluded in 2002, documented a urinary 8-oxo-7,8-dihydro-2'-deoxyguanosine level as a strong predictor of the development of diabetic nephropathy in type 2 diabetic patients (113). In addition, antioxidants were found to attenuate oxidative stress and apoptosis in human tubular cells (290).

Over the past few decades, increased production of AGE has been recognized as one of the noteworthy mechanisms that lead to diabetic complications (32, 34, 35). Studies have shown the presence of significantly increased serum and skin levels of AGE in relationship to progression of microalbuminuria to overt nephropathy and severity of renal complications in diabetic patients, respectively (22, 193).

Increased AGE production leads to enhanced free radical generation. Mechanisms responsible for AGE-induced free radical production include creation of catalytic sites for free radical generation, stimulation of NADPH oxidase, and depletion of glutathione peroxidase, a potent cellular antioxidant (297, 303). AGE-mediated mitochondrial dysfunction, in combination with carbonyl intermediates, is a source of superoxide generation, another potent free radical (238).

There is a strong synergistic correlation between oxidative stress and AGE, as indicated by the proportional increase in free radicals and AGE, in diabetic rats (265). Also, strict glycemic control has been shown to attenuate free radical production and AGE accumulation in human diabetic glomeruli (210, 266).

Neuropathy

Diabetic neuropathy represents peripheral and autonomic nerve dysfunction. It is responsible for considerable morbidity and mortality associated with diabetes. More than 50% of diabetics develop neuropathy during their lifetime, making it the most common cause of nontraumatic limb amputations (76, 293). There is a 15% chance for a patient with diabetic neuropathy of undergoing one or more amputations during his/her life (75). Additionally, poor glycemic control and duration of diabetes have been shown to be proportionally related to incidence of diabetic neuropathy (74).

As discussed earlier, hyperglycemia plays a major role in the development and progression of diabetic complications, including neuropathy. The key mechanism, implicated in the neural degeneration induced by hyperglycemia, is increased oxidative stress associated with increased polyol pathway activity, increased PKC activity, and AGE accumulation (Fig. 3). All these converge to mediate apoptosis of neurons and Schwann cells, which are the glial cells of the peripheral nervous system (242, 243, 251).

The role of oxidative stress in neuronal degeneration has been documented in multiple studies (56, 142, 213). Neurons are also vulnerable to free radical-mediated injury due to their reaction with the lipid and protein content of the neurons. This renders the neurons incapable of signaling and axonal transport with increased necrosis and apoptosis (6, 56, 189). Recent *in vivo* and *in vitro* studies have demonstrated

the role of mitochondrial dysfunction and oxidative stress as being deleterious to neurons, causing neuronal death (243, 292). Vincent *et al.* demonstrated significantly elevated levels of oxidative stress in the dorsal root ganglia, within 2 h of hyperglycemia, leading to apoptosis. This may explain in part the underlying mechanism responsible for neuropathy in diabetics with good overall control as well as patients with impaired glucose tolerance, who also develop neuropathy (225, 291). Moreover, recovery of damaged neurons in the presence of free radicals, like NO, is much slower and this recovery is accelerated by the administration of NOS inhibitors (268).

Retinopathy

Diabetic retinopathy is a severely disabling microvascular complication and is the most common diabetic eye disease (146). It is clinically manifested by multiple microvascular pathologies, including microaneurysms, hemorrhages, and neovascularization (4). Diabetic retinopathy is the most frequent cause of new cases of blindness among adults aged 20–74 years, and studies have shown that poor glucose control, per se, is the inciting factor in its development (5, 68). During the first 20 years of the disease, >60% of the patients with type 2 diabetes develop retinopathy (5). The strongest predictor for development and progression of retinopathy is the duration of diabetes itself (145).

Poor glycemic control has been consistently shown to be a key risk factor for the development of diabetic complications, including retinopathy. Moreover, tight glucose control delays the onset and progression of retinopathy in the diabetic population (272, 283). As is the case with all other diabetic complications, increased free radical production is seen in the retina of diabetics and oxidative stress is a major contributor in the pathogenesis of diabetic retinopathy (151, 208, 209) (Fig. 7).

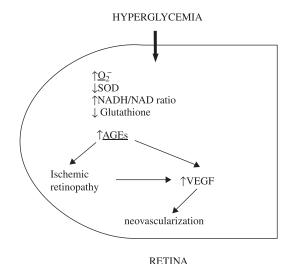


FIG. 7. Oxidative stress and pathogenesis of diabetic retinopathy. AGEs, advanced glycation end products; NAD, oxidized nicotinamide-adenine dinucleotide; NADH, nicotinamide-adenine dinucleotide; O₂⁻⁺, superoxide; SOD, superoxide dismutase; VEGF, vascular endothelial growth factor.

The retina is highly rich in polyunsaturated lipid membranes, making it extremely vulnerable to free radical-mediated lipid peroxidation (132). Thiobarbituric acid—reacting substances (TBARS) assay (126)—a tool used to measure lipid peroxide levels in oxidative stress, have been found to be increased in type 1 and type 2 diabetic patients (11, 223, 246). The vitreous fluid in diabetic patients with proliferative retinopathy has increased levels of vascular endothelial growth factor (VEGF), which is upregulated and released in response to ischemia and oxidative products (2, 3, 155, 168).

Hyperglycemia causes increased NADH-to-NAD ratio secondary to increased reduction of NAD+ to NADH. This leads to ischemic retinopathy and increased production of the superoxide (287). Moreover, superoxide dismutase (SOD) activity is decreased in the diabetic retina (65), further enhancing the superoxide activity and leading to greater oxidative stress-mediated injury and advanced glycation end (AGE) products. These essentially lead to increased release of VEGF (155, 168) and the risk of neovascularization (2, 104). The accumulation of AGE products is accelerated with advanced age and chronic hyperglycemia and is an established mechanism that mediates tissue damage, including diabetic retinopathy (250).

Further evidence implicating oxidative stress in the pathogenesis of diabetic retinopathy comes from studies showing antioxidants, such as vitamin E, normalize preclinical diabetic retinopathy (51). Additionally, increased expression of mitochondrial SOD leads to decreased superoxide formation leading to prevention of apoptosis in the retina and its capillaries (152). As in diabetic nephropathy, 8-hydroxy-2'-deoxyguanosine (8-OHdG) levels are increased in diabetic retina, and decreased levels in rats have been achieved by administration of alpha lipoic acid—an antioxidant (153). Under hyperglycemic states, glutathione a naturally occurring antioxidant, is significantly decreased in diabetic retina and this decrease is inhibited by administration of antioxidants (153, 196).

Oxidative stress also significantly contributes to the pathogenesis of diabetic cataract, another manifestation of long-standing diabetes (48, 121). The major contributors in the genesis of free radicals in diabetic lens are glycoxidation (9, 301) and impaired antioxidant defense mechanism, resulting from depletion of glutathione reserves (88, 167). Moreover, studies have documented the role of aldose reductase and sorbitol dehydrogenase, enzymes involved in polyol pathway, in the pathogenesis of slowly evolving diabetic cataract. This is thought to occur due to dysequilibrium between free radicals and antioxidant defenses resulting from depletion of NADPH & NAD+, cofactors for aldose reductase and sorbitol dehydrogenase, respectively (277, 300). In short, one of the key mechanisms in the generation of oxidative stress in diabetic cataract is the polyol pathway.

OXIDATIVE STRESS AND PREVENTION OF TYPE 2 DIABETES

In the face of the rapidly growing diabetes epidemic, which is generally perceived as a difficult-to-control disease, and with accumulating data indicating suboptimal control of cardiovascular risk factors in diabetes (182, 249), the need for orchestrated efforts to prevent diabetes has become apparent. In the

past two decades, there have been several major trials designed and executed with diabetes prevention as the primary outcome (184) (Table 1). Other studies on diabetes prevention are also ongoing (Table 1). However, it is important to note that the concept of diabetes prevention was first entertained by a pioneer scholar Elliot Joslin as early as the 1920s (131).

Among the major interventions that were tested in several well-designed randomized controlled trials, lifestyle modifications such as diet and exercise consistently yielded significant results. The Swedish Malmo feasibility study showed that men randomized to diet and exercise had less than half the risk of developing diabetes in 6 years (69). The United States Diabetes Prevention Program (DPP) enrolled >3,000 adults with impaired glucose tolerance and studied the effects of standard versus intensive lifestyle intervention, with and without metformin. Although both metformin and lifestyle interventions significantly reduced the incidence of diabetes, this effect was much more pronounced with intensive lifestyle intervention (58). Similarly, multiple international studies, in various cultural, ethnic, and social groups, have shown the benefits of lifestyle modifications for the prevention of type 2 diabetes (150, 214, 282).

Several pharmacological agents have been examined in prospective clinical trials for diabetes prevention in patients with impaired glucose tolerance. Collectively these agents inhibit reactive oxygen species through various mechanisms (Fig. 8). These include metformin, acarbose, thiazolidine-diones (TZDs), and angiotensin converting enzyme (ACE) inhibitors, among others. The preventive effects of these therapeutic modalities are primarily attributed to their direct effects on glucose lowering, reduction in triglycerides, and inhibition of the renin angiotensin–aldosterone system (RAAS).

Although lifestyle changes, metformin, acarbose, and TZDs have direct glucose-lowering effects, which may translate into prevention of diabetes and its complications, the fact that diabetes is reduced in patients on statins and ACE inhibitors, albeit inconsistently, raises the possibility of other mechanisms at work. Most of the agents that have been, so far, proven to reduce the incidence of diabetes, are also known to reduce oxidative stress. As discussed earlier in detail, oxidative stress plays a major role in the etiology, pathogenesis, and progression of diabetes and its complications.

One of the most extensively studied strategies for diabetes prevention includes lifestyle interventions such as dietary modification and exercise. These hygienic measures which invariably show improved insulin sensitivity (233) are also known to significantly decrease the levels of oxidative stress (Fig. 8), as measured by 8-isoprostaglandin F2a, superoxide, and hydrogen peroxide production, as well as improve endothelial function in diabetics (16, 173, 235, 274). Moreover, in diabetic patients, the effect of short-term lifestyle interventions has been shown to be long lasting, persisting over a 2- to 3-year follow up (234).

As discussed earlier, hyperglycemia is known to cause enhanced protein kinase C (PKC) activity (33, 124) and metformin, one of the most widely used oral hypoglycemics, not only lowers glucose, but also reduces oxidative stress by inhibiting PKC (143) (Fig. 8). Additionally, metformin reduces oxidative stress-mediated injury by scavenging free radicals and blunting plasma membrane NADPH oxidase—another feature of chronic hyperglycemia (30, 83, 143).

TABLE 1. DIABETES PREVENTION TRIALS

Intervention	Trial	Population (number)	NNT
Completed trials with di	abetes prevention as primary outcome		
Lifestyle changes	DPP	IFG/IGT ($n = 3,234$)	7
, E	FDPS	IGT(n = 522)	8
Metformin	DPP	IFG/IGT $(n = 3,234)$	14
Thiazolidinediones	DREAM (Roziglotazone)	IFG/IGT $(n = 5,269)$	7
Acarbose	STOP-NIDDM	IFG/IGT (n = 1,429)	11
ACE-inhibitors	DREAM (ramipril)	1FG/IGT (n = 5,269)	NS
Xenical	XENDOS	All Obese $(n = 3,305)$	36
		Obese + IGT $(n = 694)$	10

Ongoing trials wth diabetes prevention as primary outcome

Nateglinide NAVIGATOR

Thiazolidinediones ACT NOW (Pioglitazone)
ARBs NAVIGATOR (Valsartan))

ONTARGET/TRANSCEND (Telmisartan)

Metformin + Rosiglotazone CANOE

Other trials*

ACE-inhibitors HOPE (ramipril), CAPP (captopril), ALLHAT (lisinopril)

ARBs LIFE (losartan)
Statins WOSCOPS
Bezafibrate BIP
Rimobanant RIO trials

Trials: ACT NOW: ACTos NOW for Prevention of Diabetes; ALLHAT: The Antihypertensive and Lipid-Lowering Treatment to Prevent Heart Attack Trial; BIP: Bezfibrate Infarction Prevention Study; CAnadian Normoglycemia Outcomes Evaluation (CANOE) trial; CAPP: The CAptopril Prevention Project; DPP: Diabetes Prevention Program; DREAM: Diabetes REduction Assessment with ramipril and roziglotazone Medications; FDPS: Finnish Diabetes Prevention Study; HOPE: Heart Outcomes Prevention Study; LIFE: Losartan Intervention For Endpoint; NAVIGATOR: Nateglinide And Valsartan in Impaired Glucose Tolerance Outcomes Research; ONTARGET: ONgoing Telmisartan Alone and in combination with Ramipril Global Endpoint Trial; RIO: Rimonabant In Obesity; STOP-NIDDM: Study TO Prevent Non-Insulin Dependent Diabetes; TRANSCEND: Telmisartan Randomized Assessment Study in ACE Intolerant Subjects with Cardiovascular disease; WOSCOPS: West of Scotland Coronary Prevention Study; XENDOS: XENical in the prevention of Diabetes in Obese Subjects.

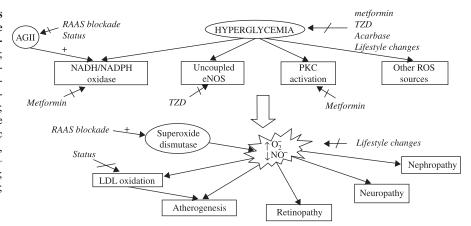
Thiazolidinedione (TZD) is a class of medication that reduces insulin resistance by reducing plasma glucose, glucose production, and increasing glucose clearance in type 2 diabetes. These agents work primarily by binding to nuclear peroxisome proliferator-activated receptor gamma (PPAR γ) (179, 264). TZDs mediate their antioxidant effects by inhibiting nitric oxide synthase, thereby decreasing peroxynitrite production, and inhibiting superoxide production (185, 231) (Fig. 8). The preventive effects of TZDs were recently validated by the rosiglitazone limb of the Diabetes Reduction Assessment with ramipril and rosiglitazone Medication (DREAM) trial (63) which showed a 60% reduction in the incidence of diabetes in patients with impaired fasting glucose or impaired glucose tolerance. Moreover, there are multiple animal studies showing additional antioxidant properties of TZDs, and researchers are exploring TZD analogues with promising results in ameliorating oxidative stress associated with diabetes (47).

ACE inhibitors have long been known for their cardioprotective and renoprotective effects, but their potential for prevention of diabetes has been a matter of debate. Although post-hoc analysis of the Heart Outcomes Prevention Evaluation (HOPE) trial (304) provided the most persuasive evidence for diabetes prevention with the use of ACE inhibitors, this has not been duplicated in the largest diabetes prevention trial to date: the DREAM trial (273). Even though ramipril, an ACE inhibitor, has not been shown to prevent diabetes, the beneficial effects of these agents on glucose metabolism were confirmed in the DREAM study by a reduction of 6 mg/dl in the 2-h post oral glucose tolerance test plasma glucose level, and by the increased conversion of prediabetes to normoglycemia by 16%. These agents that block RAAS have well-proven effects on the reduction of oxidative stress (26, 115).

Angiotensin II increases intracellular superoxide by the activation of NADPH and NADH oxidase, resulting in oxidative stress (92). This plays a significant role in insulin resistance, and animal studies suggest that abrogation of oxidative stress improves insulin sensitivity (26). Furthermore, it also activates NADPH by enhancing the gene expression of GTPase Rac-1, leading to increased production of peroxynitrite (296). In addition to inhibiting the production of angiotensin II, ACE inhibitors increase the activity of endothelial superoxide

^{*}Trials with diabetes prevention as secondary outcome or on posthoc analysis. ACE, angiotensin converting enzyme; ARBs, angiotensin receptor blockers; GD: previous gestational diabetes; IFG, impaired fasting glucose; IGT, impaired glucose tolerance; NNT, number needed to treat; NS, not significant.

FIG. 8. Effects of diabetes prevention agents on oxidative stress associated with hyperglycemia. AGII, angiotensin II; eNOS, endothelial nitric oxide synthase; LDL, low-density lipoprotein, NADP, oxidized nicotinamideadenine dinucleotide phosphate; NADPH, nicotinamide-adenine dinucleotide phosphate; NO, nitric oxide; O₂-, superoxide; PKC, protein kinase C; RAAS, reninangiotensin-aldosterone system; ROS, reactive oxygen species; TZD, thiazolidinediones.



dismutase, a major antioxidant (115) (Fig. 8). Other agents that inhibit the RAAS include angiotensin receptors blockers, which are currently being tested in the Nateglinide And Valsartan in Impaired Glucose Tolerance Outcomes Research (NAVIGATOR) trial for primary diabetes prevention (Table 1).

Finally, among other agents that have shown favorable effects on glucose metabolism, statins (Table 1) also reduce oxidative stress independent of LDL reduction (66, 183) (Fig. 8). This reduction in oxidative stress is mediated by various mechanisms such as attenuation of angiotensin II-mediated free radical generation (288) and reduction of LDL oxidation (180).

CONCLUSION

In this article, we have presented a comprehensive review highlighting the evidence that oxidative stress serves as a common soil hypothesis for the various interventions that have been examined in large prospective randomized trials with prevention of diabetes being the primary outcome. These interventions included lifestyle changes as well as pharmacological agents, such as metformin, acarbose, and TZDs, that have been shown to be as effective as lifestyle interventions in the prevention of diabetes, as indicated in the recently published DREAM trial, the largest diabetes prevention trial to date.

The prospect for the use of antioxidants specifically for the prevention of diabetes holds a great promise in the continuing efforts to curb the rapidly growing diabetes epidemic. However, further studies are needed to provide specific answers in this important area of research.

ACKNOWLEDGMENT

This work was supported by grant from the American Diabetes Association, 7-05-RA-89, to SIM.

ABBREVIATIONS

ACE, angiotensin converting enzyme; ADA, American Diabetes Association, ADMA, asymmetric dimethylarginine; AGE, advanced glycation end products; AIDS, acquired

immunodeficiency syndrome; ARDS, adult respiratory distress syndrome; ATP, adenosine triphosphate; 13C, carbon isotope; CAD, coronary artery disease; CHD, coronary heart disease; CHF, congestive heart failure; CMS, cardiometabolic syndrome; DNA, deoxyribonucleic acid; DPP, Diabetes Prevention Program; DREAM, Diabetes Reduction Assessment with Ramipril and Rosiglitazone Medication; eNOS, endothelial nitric oxide synthase; ESR, electron spin resonance; FFA, free fatty acids; GFR, glomerular filtration rate; GLUT-4, glucose transporter isoform-4; GTP, guanosine triphosphate; H₂O₂, hydrogen peroxide; HDL, high density lipoprotein; HOPE, Heart Outcomes Prevention Evaluation; 'OH. Hydroxyl; 8-OHdG, 8-hydroxy-2'-deoxyguanosine; IUGR, Intrauterine growth retardation; LDL, low density lipoprotein; MCP-1, monocyte chemoattractant protein; MDA, malondialdehyde: NAD+, nicotinamide adenine dinucleotide (oxidized form); NADH, nicotinamide adenine dinucleotide (reduced form); NADPH, nicotinamide adenine dinucleotide phosphate; NAVIGATOR, Nateglinide And Valsartan in Impaired Glucose Tolerance Outcomes Research; NCEP/ATP, National Cholesterol Education Program/Adult Treatment Panel; NO, nitric oxide; NO, nitronium ion; NO, nitrogen dioxide gas; NOS, nitric oxide synthase; O2-, superoxide; ONOO-, peroxynitrite; ORIGIN, Outcomes Reduction with Initial Glargine Intervention; oxLDL, oxidized low density lipoprotein; Ph. C*, triphenylmethyl radical; 31P, phosphorus isotope; PKC, protein kinase C; PPAR y, peroxisome proliferator-activated receptor gamma; RAAS, renin angiotensin aldosterone system; RO', alkoxyl; ROO', peroxyl; ROS, reactive oxygen species; RS', thiyl; SOD, superoxide dismutase; TBARS, thiobarbituric acid-reactive substances; TZD, thiazolidinedione; VEGF, vascular endothelial growth factor; VSMC, vascular smooth muscle cells; WHO, World Health Organization.

REFERENCES

- Abbott RD, Kannel RP, and Wilson WB. The impact of diabetes on survival following myocardial infarction in men vs women: the Framingham Study. *JAMA* 260: 3456–3460, 1988.
- Adamis AP, Miller JW, Bernal MT, D'Amico DJ, Folkman J, Yeo TK, and Yeo KT. Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. Am J Ophthalmol 118: 445–450, 1994.

- Aiello LP, Avery RL, Arrigg PG, Keyt BA, Jampel HD, Shah ST, Pasquale LR, Thieme H, Iwamoto MA, Park JE, Nguyen HV, Aiello LM, Ferrara N, and King GL. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. N Engl J Med 331: 1480–1487, 1994.
- Aiello LP, Cahill M, and Wong JS. Systemic considerations in the management of diabetic retinopathy. Am J Ophthalmol 132: 760–776, 2001.
- Aiello LP, Gardner TW, King GL, Blankenship G, Cavallerano JD, Ferris FL III, and Klein R. Diabetic retinopathy (Technical Review). *Diabetes Care* 21: 143–156, 1998.
- Aksenova MV, Aksenov MY, Payne RM, Trojanowski JQ, Schmidt ML, Carney JM, Butterfield DA, and Markesbery WR. Oxidation of cytosolic proteins & expression of creatine kinase BB in frontal lobe in different neurodegenerative disorders. Dement Geriatr Cogn Disord 10: 158–165, 1999.
- Alderton WK, Cooper CE, and Knowles RG. Nitric oxide synthases: structure, function and inhibition. *Biochem J* 357: 593–615, 2001.
- Ames BN and Shigenage MK. Oxidants are a major contributor to cancer and aging. In: *DNA and Free Radicals*, edited by Halliwell B, Aruoma O. New York: Ellis Harwood Ltd, 1993, pp. 3–15.
- Ansari NH, Wang L, Erwin AA, and Church DF. Glucose-dependent formation of free radical species in lens homogenate. *Biochem Mol Med* 59: 68–71, 1996.
- Antinozzi PA, Ishihara H, Newgard CB, and Wollheim CB. Mitochondrial metabolism sets the maximal limit of fuel-stimulated insulin secretion in a model pancreatic beta cell. A survey of four fuel secretagogues. *J Biol Chem* 277: 11746–11755, 2002.
- Armstrong D, Abdella N, Salman A, Miller N, Rahman EA, and Bojancyzk M. Relationship of lipid peroxides to diabetic complications: comparison with conventional laboratory tests. *J Diabetes Complications* 6: 116–122, 1992.
- Assmann G, Carmena R, Cullen P, Fruchart JC, Jossa F, Lewis B, Mancini M, and Paoletti R. Coronary heart disease: reducing the risk: a worldwide view. International Task Force for the Prevention of Coronary Heart Disease. *Circulation* 100: 1930–1938, 1999.
- Autiero M, Sansone G, and Abrescia P. Relative ratios of lactoferrin, albumin, and acid phosphatase seminal levels as sperm quality markers in fertile and infertile men. *J Androl* 12: 191–200, 1991.
- Bae JH, Bassenge E, Kim KB, Kim YN, Kim KS, Lee HJ, Moon KC, Lee MS, Park KY, and Schwemmer M. Postprandial hypertriglyceridemia impairs endothelial function by enhanced oxidant stress. *Atherosclerosis* 155: 517–523, 2001.
- 15. Baker K, Marcus CB, Huffman K, Kruk H, Malfroy B, and Doctrow SR. Synthetic combined superoxide dismutase/catalase mimetics are protective as a delayed treatment in a rat stroke model: a key role for reactive oxygen species in ischemic brain injury. J Pharmacol Exp Ther 284: 215–221, 1998.
- Baldus S, Heitzer T, Eiserich JP, Lau D, Mollnau H, Ortak M, Petri S, GOldmann B, Duchstein HJ, Berger J, Helmchen U, Freeman BA, Meinertz T, and Munzel R. Myeloperoxidase enhances nitric oxide catabolism during myocardial ischemia and reperfusion. Free Radic Biol Med 37: 902–911, 2004.
- Bavdekar A, Sachdev HS, Fall CHD, Osmond C, Lakshmy R, Barker DJP, Biswas SKD, Ramji S, Prabhakaran D, and Reddy KS. Relation of serial changes in childhood body-mass index to impaired glucose tolerance in young adulthood. N Engl J Med 350: 865–875, 2004.
- Baynes JW. Role of oxidative stress in development of complications in diabetes. *Diabetes* 40: 405–412, 1991.
- Beckman JA, Goldfine AB, Gordon MB, and Creager MA. Ascorbate restores endothelium-dependent vasodilation impaired by acute hyperglycemia in humans. *Circulation* 103: 1618–1623, 2001.
- Beckman JS, Chen J, Ischiropoulos H, and Crow JP. Oxidative chemistry of peroxynitrite. Meth Enzymol 233: 229–240, 1994.
- Beckman KB and Ames BN. Endogenous oxidative damage of mtDNA. Mutat Res 424: 51–58, 1999.

 Beisswenger PJ, Drummond KS, Nelson RG, Howell SK, Szwergold BS, and Mauer M. Susceptibility to diabetic nephropathy is related to dicarbonyl and oxidative stress. *Diabetes* 54: 3274–3281, 2005.

- Bergendi L, Benes L, Durackova Z, and Ferencik M. Chemistry, physiology and pathology of free radicals. *Life Sci* 65: 1865–1874, 1999.
- 24. Betteridge DJ. What is oxidative stress? (Review). *Metabolism* 49: 3–8, 2000.
- Biasi F, Bosco M, Chiappino I, Lanfranco G, Ottobrelli A, Massano G, Donadio PP, Vaj M, Adoreno E, et al. Oxidative damage in human liver transplantation. Free Radic Biol Med 19: 311–317, 1995.
- 26. Blendea MC, Jacobs D, Stump CS, McFarlane SI, Ogrin C, Bahtyiar G, Stas S, Kumar P, Sha Q, Ferrario CM, and Sowers JR. Abrogation of oxidative stress improves insulin sensitivity in the Ren-2 rat model of tissue angiotensin II overexpression. Am J Physiol Endocrinol Metab 288: E 353–359, 2005.
- 27. Blendea MC, McFarlane SI, Isenovic ER, Gick G, and Sowers JR. Heart disease in diabetic patients. *Curr Diab Rep* 3: 223–229, 2003.
- Block G, Dietrich M, Norkus EP, Morrow JD, Hudes M, Caan B, and Packer L. Factors associated with oxidative stress in human populations. *Am J Epidemiol* 156: 274–285, 2002.
- Bohr VA and Dianov GL. Oxidative DNA damage processing in nuclear and mitochondrial DNA. *Biochimie* 81: 155–160, 1999.
- Bonnefont–Rousselot D, Raji B, Walrand S, Gardes–Albert M, Jore D, Legrand A, Peynet J, and Vasson MP. An intracellular modulation of free radical production could contribute to the beneficial effects of metformin towards oxidative stress. *Metabolism* 5: 586–589, 2003.
- Booth GL, Kapral MK, Fung K, and Tu JV. Relation between age and cardiovascular disease in men and women with diabetes compared with non-diabetic people: a population-based retrospective cohort study. *Lancet* 368: 29–36, 2006.
- 32. Brownlee M. Glycation products and the pathogenesis of diabetic complications. *Diabetes Care* 15: 1835–1843, 1992.
- Brownlee M. Biochemistry and molecular cell biology of diabetic complications. *Nature* 414: 813–820, 2001.
- Bucala R and Cerami A. Advanced glycosylation: chemistry, biology, and implications for diabetes and aging (Review). Adv Pharmacol 23: 1–34, 1992.
- Bunn HF and Higgins PJ. Reaction of monosaccharides with proteins: possible evolutionary significance. *Science* 213: 222–224, 1981.
- Cai H and Harrison DG. Endothelial dysfunction in cardiovascular disease: the role of oxidant stress. Circ Res 87: 840–844, 2000
- Cai L and Kang YJ. Cell death and diabetic cardiomyopathy. Cardiovasc Toxicol 3: 219–228, 2003.
- Cai L, Li W, Wang G, Guo L, Jiang Y, and Kang YJ. Hyperglycemia-induced apoptosis in mouse myocardium: mitochondrial cytochrome C-mediated caspase-3 activation pathway. *Diabetes* 51: 1938–1948, 2002.
- Caramori MLA and Mauer M. Diabetes and nephropathy. Current Opinion Nephrol Hypertension 12: 273–282, 2003.
- Carpagnano GE, Kharitonov A, Resta O, Foschino–Barbaro MP, Gramiccioni E, and Barnes PJ. 8-Isoprostane, a marker of oxidative stress, is increased in exhaled breath condensate of patients with obstructive sleep apnea after night and is reduced by continuous positive airway pressure therapy. *Chest* 124: 1386–1392, 2003
- Center for Disease Control (CDC). National Diabetes Fact Sheet, 2005. (Accessed 12/7/2006, at www.cdc.gov/diabetes/ pubs/pdf/ndfs 2005.pdf)
- Center for Disease Control (CDC). National Diabetes Surveillance System. (Accessed 12/7/2006, at http://www.cdc.gov/diabetes/statistics.)
- Centers for Disease Control and Prevention. National diabetes fact sheet: general information and national estimates on diabetes in the United States, 2003. Atlanta, GA: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, 2004.

- 44. Ceriello A, Motz E, and Giugliano D. Vitamin C and hypertension. *Lancet* 355: 1271–1272, 2000.
- Cerutti PA. Oxidant stress and carcinogenesis. Europ J Clin Invest 21: 1–5, 1991.
- Chandel NS, Budinger GRS, and Schumacker PT. Molecular oxygen modulates cytochrome c oxidase function. *J Biol Chem* 271: 8672–8677, 1996.
- Chaudhry J, Ghosh NN, Roy K, and Chandra R. Antihyperglycemic effect of a new thiazolidinedione analogue and its role in ameliorating oxidative stress in alloxan-induced diabetic rats. *Life Sci* 80: 1135–1142, 2007.
- Cheng HM and Gonzalez RG. The effect of high glucose and oxidative stress on lens metabolism, aldose reductase, and senile cataractogenesis. *Metabolism* 35: 10–14, 1986.
- Chiarelli F, Cipollone F, Romano F, Tumini S, Costantini F, di Ricco L, Pomilio M, Pierdomenico SD, Marini M, Cuccurullo F, and Mezzetti A. Increased circulating nitric oxide in young patients with type 1 diabetes and persistent microalbuminuria: relation to glomerular hyperfiltration. *Diabetes* 49: 1258–1263, 2000.
- Chiueh CC. Neuroprotective properties of nitric oxide. Ann NY Acad Sci 890: 301–311, 1999.
- Clermont AC, Aiello LP, Aiello LM, Schlossman D, Kopple A, King GL, and Burcell SE. Vitamin E normalized retinal blood flow in diabetic patients with minimal diabetic retinopathy: results of a double masked crossover clinical trial (Abstract). *Invest Ophthalmol Vis Sci 39* (Suppl. 4): S1000, 1998.
- Cohen G. Oxy-radical toxicity in catecholamine neurons. Neurotoxicology 5: 77–82, 1984.
- Dandona P, Aljada A, Chaudhuri A, Mohanty P, and Garg R. A comprehensive perspective based on interactions between obesity, diabetes, and inflammation. *Circulation* 111: 1448–1454, 2005.
- 54. Dandona P, Mohanty P, Ghanim H, Aljada A, Browne R, Hamouda W, Prabhala A, Afzal A, and Garg R. The suppressive effect of dietary restriction and weight loss in the obese on the generation of reactive oxygen species by leukocytes, lipid peroxidation, and protein carbonylation. *J Clin Endocrinol Metab* 86: 355–362, 2001.
- Davies GR, Simmonds NJ, Stevens TRJ, Grandison A, Blake DR, and Rampton DS. Mucosal reactive oxygen metabolite production in duodenal ulcer disease. *Gut* 33: 1467–1472, 1992.
- Deng G, Su JH, Ivins KJ, Van Houten B, and Cotman CW. Bcl-2 facilitates recovery from DNA damage after oxidative stress. *Exp Neurol* 159: 309–318, 1999.
- 57. Dhindsa S, Tripathy D, Mohanty P, Ghanim H, Syed T, Aljada A, and Dandona P. Differential effects of glucose and alcohol on reactive oxygen species generation and intranuclear nuclear factor-kappaB in mononuclear cells. *Metab Clin Exp* 53: 330–334, 2004.
- Diabetes Prevention Program Research Group. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. N Engl J Med 346: 393–403, 2002.
- Diederich D, Yang ZH, Buhler FR, and Luscher TF. Impaired endothelium-dependent relaxations in hypertensive resistance arteries involve cyclooxygenase pathway. Am J Physiol Heart Circ Physiol 258: H445–H451, 1990.
- Diederich D, Skopec J, Diederich A, and Dai F. Endothelial dysfunction in mesenteric resistance arteries of diabetic rats: role of free radicals. *Am J Physiol* 266: H1153–H1161, 1994.
- Dizdaroglu M. Chemistry of free radical damage to DNA and nucleoproteins. In: DNA and Free Radicals. Halliwell B, Aruoma Ol, eds. Chichester: Ellis Harwood, 1993, pp. 19–39.
- Donath MY, Gross DJ, Cerasi E, and Kaiser N. Hyperglycemiainduced β-cell apoptosis in pancreatic islets of *Psammomys obesus* during development of diabetes. *Diabetes* 48: 738–744, 1999.
- 63. DREAM (Diabetes Reduction Assessment with ramipril and rosiglitazone Medication) Trial Investigators. Effect of rosiglitazone on the frequency of diabetes in patients with impaired glucose tolerance or impaired fasting glucose: a randomized controlled trial. *Lancet* 368: 1096–105, 2006.
- 64. Du XL, Edelstein D, Rossetti L, Fantus IG, Goldberg H, Ziyadeh F, Wu J, and Brownlee M. Hyperglycemia-induced mitochondrial superoxide overproduction activates the hexosamine pathway and induces plasminogen activator inhibitor-1 expression by

- increasing Sp1 glycosylation. *Proc Natl Acad Sci USA* 97: 12222–12226, 2002.
- Du Y, Miller CM, and Kern TS. Hyperglycemia increases mitochondrial superoxide in retina and retinal cells. *Free Radic Biol Med* 35: 1491–1499, 2003.
- 66. Endo K, Miyashita Y, Sasaki H, Ebisuno M, Obira M, Saiki A, Koide N, Oyama T, Takeyoshi M, and Shirai K. Probucol and atorvastatin decrease urinary 8-hyrdoxy-2-deoxyguanosine in patients with diabetes and hypercholesterolemia. *J Atheroscler Thromb* 13: 68–75, 2006.
- Engerman RL, Kern TS, and Larson ME. Nerve conduction and aldose reductase inhibition during 5 years of diabetes or galactosaemia in dogs. *Diabetologia* 37: 141–144, 1994.
- Engerman RL and Kern TS. Experimental galactosemia produces diabetic-like retinopathy. *Diabetes* 33: 97–100, 1984.
- 69. Eriksson KF and Lindgarde F. Prevention of type 2 (non-insulin-dependent) diabetes mellitus by diet and physical exercise: the 6-year Malmo feasibility study. *Diabetologia* 34: 891–898, 1991.
- Esposti MD and McLennan H. Mitochondria and cells produce reactive oxygen species in virtual anaerobiosis: relevance to ceramide-induced apoptosis. FEBS Lett 430: 338–342, 1991.
- 71. Evans JL, Goldfine ID, Maddux BA, and Grodsky GM. Are oxidative stress-activated signaling pathways mediators of insulin resistance and h-cell dysfunction? *Diabetes* 52: 1–8, 2003.
- Evans JL, Goldfine ID, Maddux BA, and Grodsky GM. Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. *Endocr Rev* 23: 599–622, 2002.
- 73. Fein FS. Diabetic cardiomyopathy. *Diabetes Care* 13: 1169–1179, 1990.
- Feldman EL, Stevens MJ, and Russell JW. Diabetic peripheral and autonomic neuropathy. In: *Contemporary Endocrinology*, edited by Sperling MA. Totowa, NJ: Humana Press. 1998, pp. 437–461
- Feldman, EL, Stevens MJ, Russell JW, and Greene DA. Diabetic neuropathy. In: *Principles and Practice of Endocrinology and Metabolism*, edited by Becker KL. Philadelphia, Lippincott Williams & Wilkins. 2001, pp. 1391–1399.
- Feldman, EL, Stevens MJ, Russell JW, and Greene DA. Somatosensory neuropathy. In: *Ellenberg and Rifkin's Diabetes Mellitus*, edited by Porte D Jr, Sherwin RS, and Baron A. New York, McGraw Hill. 2002, pp. 771–788.
- 77. Fenton HJH. Oxidation of tartaric acid in presence of iron. *J Chem Soc* 65: 889–910, 1894.
- Flores SC, Marecki JC, Harper KP, Bose SK, Nelson SK, and McCord JM. Tat protein of human immunodeficiency virus type 1 represses expression of manganese superoxide dismutase in HeLa cells. *Proc Natl Acad Sci USA* 90: 7632–7636, 1993.
- Folsom AR, Rasmussen ML, Chambless LE, Howard G, Cooper LS, Schmidt MI, and Heiss G. Prospective associations of fasting insulin, body fat distribution, and diabetes with risk of ischemic stroke. The Atherosclerosis Risk in Communities (ARIC) Study Investigators. *Diabetes Care* 22: 1077–1083, 1000
- Forsen T, Eriksson J, Tuomilehto J, Reunanen A, Osmond C, and Barker D. The fetal and childhood growth of persons who develop type 2 diabetes. *Ann Intern Med* 133: 176–182, 2000.
- 81. Fukumoto H, Naito Z, Asano G, and Aramaki T. Immunohistochemical and morphometric evaluations of coronary atherosclerotic plaques associated with myocardial infarction and diabetes mellitus. *J Atheroscler Thromb* 5: 29–35, 1998.
- Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, Nakayama O, Makishima M, Matsuda M, and Shimomura I. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 114: 1752–1761, 2004.
- 83. Gallo A, Ceolotto G, Pinton P, Iori E, Murphy E, Rutter GA, Rizzuto R, Semplicini A, and Avogaro A. Metformin prevents glucose-induced protein kinase c-β2 activation in human umbilical vein endothelial cells through an antioxidant mechanism. *Diabetes* 54: 1123–1131, 2005.
- 84. Gavras I and Gavras H. Angiotensin II as a cardiovascular risk factor. *J Hum Hypertens* 16: S2–S6, 2002.

 Giugliano D, Ceriello A, and Paolisso G. Oxidative stress and diabetic vascular complications. *Diabetes Care* 19: 257–267, 1996.

- Glavind J, Hartman S, Clemmensen J, Jessen KE, and Dam H. Studies on the role of lipoperoxidase in human pathology: the presence of peroxide lipids in atherosclerotic aorta. *Acta Pathol Microbiol Scand* 30: 1–6, 1952.
- 87. Gomberg M. An instance of trivalent carbon: triphenylmethyl. JAm Chem Soc 22: 757–71, 1900.
- 88. Gonzalez AM, Sochor M, and McLean P. The effect of an aldose reductase inhibitor (Sorbinil) on the level of metabolites in lenses of diabetic rats. *Diabetes* 32: 482–485, 1983.
- Gonzalez PK, Zhuang J, Doctrow SR, Malfroy B, Benson PF, Menconi MJ, and Fink MP. Role of oxidant stress in the adult respiratory distress syndrome: evaluation of a novel antioxidant strategy in a porcine model of endotoxin-induced acute lung injury. *Shock* 1(6 suppl): S23–S26, 1996.
- Grankvist K, Marklund SL, and Taljedal IB. CuZn-superoxide dismutase, Mn-superoxide dismutase, catalase and glutathione peroxidase in pancreatic islets and other tissues in the mouse *Biochem J* 199: 393–398, 1981.
- Greene DA, Sima AA, Stevens MJ, Feldman EL, and Lattimer SA. Complications: neuropathy, pathogenetic considerations. *Diabetes Care* 15: 1902–1925, 1992.
- Griendling KK, Sorescu D, Lassegue B, and Ushio-Fukai M. Modulation of protein kinase activity and gene expression by reactive oxygen species and their role in vascular physiology and pathophysiology. *Arterioscler Thromb Vasc Biol* 20: 2175–2183, 2000.
- 93. Grundy, SM, Cleeman, JI, Daniels, SR, Donato KA, Eckel RH, Franklin BA, Gordon DJ, Krauss RM, Savage PJ, Smith SC Jr, Spertus JA, and Costa F. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute Scientific Statement. *Circulation* 112: 2735, 2005.
- 94. Guo Q, Qian SY, and Mason RP. Separation and identification of DMPO adducts of oxygen-centered radicals formed from organic hydroperoxides by HPLC-ESR, ESI-MS and MS/MS. *J Am Soc Mass Spectrom* 14: 862–871, 2003.
- Guzik TJ, Mussa S, Gastaldi D, Sadowski J, Ratnatunga C, Pillai R, and Channon KM. Mechanisms of increased vascular superoxide production in human diabetes mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase. *Circulation* 105: 1656–1662, 2002.
- Ha H, Park J, Kim YS, and Endou H. Oxidative stress and chronic allograft nephropathy. (Review). *Yonsei Med J* 45: 1049–1052, 2004.
- Haber CA, Lam TK, Yu Z, Gupta N, Goh T, Bogdanovic E, Giacca A, and Fantus IG. N-acetylcysteine and taurine prevent hyperglycemia-induced insulin resistance in vivo: possible role of oxidative stress. *Am J Physiol Endocrinol Metab* 285: E744–E753, 2003.
- 98. Haffner S, D'Agostino RJ, Saad M, O'Leary D, Savage P, Rewers M, Selby J, Bergman R, and Mykkanen L. Carotid artery atherosclerosis in type-2 diabetic and nondiabetic subjects with and without symptomatic coronary artery disease: the Insulin Resistance Atherosclerosis Study. Am J Cardiol 85: 1395–1400, 2000.
- 99. Haffner S, Lehto S, Ronnemaa T, Pyorala K, and Laakso M. Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. N Engl J Med 339: 229–234, 1998.
- Hall DB, Holmin RE, and Barton JK. Oxidative DNA damage through long range electron transfer. *Nature* 382: 731–735, 1996.
- Halliwell B and Gutteridge JMC. Free Radicals in Biology and Medicine. 2nd ed. Oxford: Clarendon Press. 1989.
- Halliwell B and Gutteridge JMC. Oxygen toxicity, oxygen radicals, transition metals and disease. *Biochem J* 219: 1–14, 1984.
- Halliwell B. Free radicals, antioxidants and human disease: Curiosity, cause or consequence. *Lancet* 344: 721–724, 1994.
- 104. Hammes HP, Weiss A, Hess S, Araki N, Horiuchi S, Brownlee M, and Preissner KT. Modification of vitronectin by advanced glycation alters functional properties *in vitro* and the diabetic retina. *Lab Invest* 75: 325–338, 1996.

105. Hansel B, Giral P, Nobecourt E, Chantepie S, Bruckert E, Chapman MJ, and Kontush A. Metabolic syndrome is associated with elevated oxidative stress and dysfunctional dense high-density lipoprotein particles displaying impaired antioxidative activity. *J Clin Endocrinol Metab* 89: 4963–4971, 2004.

- Harman D. Ageing. A theory based on free radical and radiation chemistry. J Gerontol 11: 298–300, 1956.
- 107. Harman D. The ageing process. *Proc Natl Acad Sci USA* 78: 7124–7128, 1981.
- Hayden MR, Chowdhury N, Govindarajan G, Karuparthi P, Habibi J, and Sowers JR. Myocardial myocyte remodeling and fibrosis in cardiometabolic syndrome. *J Cariometab Syndr* 1: 326–333, 2006.
- Hayden MR, Stump CS, and Sowers JR. Introduction: organ involvement in cardiometabolic syndrome. *J Cariometab Syndr* 1: 16–24, 2006.
- 110. Hayden MR and Tyagi SC. Myocardial redox stress and remodeling in metabolic syndrome, type 2 diabetes mellitus, and congestive heart failure. Med Sci Monit 9: SR47–64, 2003.
- 111. Hayden MR, Whaley-Connell A, and Sowers JR. Renal redox stress & remodeling in metabolic syndrome, type 2 diabetes mellitus, and diabetic nephropathy: Paying homage to the podocyte. Am J Nephrol 25: 553–569, 2005.
- Herlitz JK, Edrardsson BW, Emanuelsson N, and Hjalmarson H. Prognosis in diabetics with chest pain or other symptoms suggestive of acute myocardial infarction. *Cardiology* 80: 237–245, 1992.
- 113. Hinokio Y, Suzuki S, Hirai M, Suzuki C, Suzuki M, and Toyota T. Urinary excretion of 8-oxo-7,8-dihydro-2'-deoxyguanosine as a predictor of the development of diabetic nephropathy. *Diabetologia* 45: 877–882, 2002.
- 114. Hofmann MA, Schiekofer S, Kanitz M, Klevesath MS, Joswig M, Lee V, Morcos M, Tritschler H, Ziegler R, Wahl P, Bierhaus A, and Nawroth PP. Insufficient glycemic control increases nuclear factor-kappa B binding activity in peripheral blood mononuclear cells isolated from patients with type 1 diabetes. *Diabetes Care* 21: 1310–1316, 1998.
- 115. Hornig B, Landmesser U, and Kohler C. Comparative effect of ACE inhibition and angiotensin II type 1 receptor antagonism on bioavailability of nitric oxide in patients with coronary artery disease. *Circulation* 103: 799–805, 2001.
- Houglum K, Ramm GA, Crawford DH, Witztum JL, Powell LW, and Chojkier M. Excess iron induces hepatic oxidative stress and transforming growth factor beta1 in genetic hemochromatosis. *Hepatology* 26: 605–610, 1997.
- 117. Hsieh CC, Yen MH, Yen CH, and Lau YT. Oxidized low density lipoprotein induces apoptosis via generation of reactive oxygen species in vascular smooth muscle cells. *Cardiovasc Res* 49: 135–145, 2001.
- 118. http://www.who.int/mediacentre/factsheets/fs312/en/ (World Health Organization (WHO) Fact sheet)
- Hu F, Stampfer M, Solomon C, Willett W, and Manson J. Diabetes mellitus and mortality from all causes and coronary heart disease in women: 20 years of follow-up. *Diabetes* 49(Suppl. 1): A20, 2001
- 120. Hubel CA. Oxidative stress in the pathogenesis of preeclampsia. *Proc Soc Exp Biol Med* 222: 222–235, 1999.
- 121. Hunt JV, Jiang ZY, and Wolff SP. Formation of hydrogen peroxide by lens proteins: protein-derived hydrogen peroxide as a potential mechanism of oxidative insult to the lens. *Free Rad Biol Med* 13: 319–323, 1992.
- 122. Inoguchi T, Li P, Umeda F, Yu HY, Kakimoto M, Imamura M, Aoki T, Etoh T, Hashimoto T, Naruse M, Sano H, Utsumi H, and Nawata H. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 49: 1939–1945, 2000.
- 123. Inoguchi T, Sonta T, Tsubouchi H, Etoh T, Kakimoto M, Sonoda N, Sato N, Sekiguchi N, Kobayashi K, Sumimoto H, Utsumi H, and Nawata H. Protein kinase C-dependent increase in reactive oxygen species (ROS) production in vascular tissues of diabetes: role of vascular NAD(P)H oxidase. *J Am Soc Nephrol* 14 (8 suppl.3): S227–S232, 2003.

- 124. Ishii H, Koya D, and King GL. Protein kinase C activation and its role in the development of vascular complications in diabetes mellitus. *J Mol Med* 76: 21–31, 1998.
- 125. Jamrozik K, Broadhurst RJ, Forbes S, Hankey GJ, and Anderson CS. Predictors of death and vascular events in the elderly: the Perth Community Stroke Study. Stroke 31: 863–868, 2000.
- 126. Janero DR. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. (Review). Free Radic Biol Med 9: 515–540, 1990.
- 127. Jarvisalo MJ, Raitakari M, Toikka J, Putto-Laurila A, Rontu R, Laine S, Lehtimaki T, Ronnemaa T, Viikari J, and Raitakari OT. Endothelial dysfunction and increased arterial intima-media thickness in children with type 1 diabetes. *Circulation* 109: 1750–1755, 2004.
- Jay D, Hitomi H, and Griendling KK. Oxidative stress and diabetic cardiovascular complication. Free Rad Biol Med 40: 183–192, 2006.
- Jonas JC, Sharma A, Hasenkamp W, Ilkova H, Patane G, Laybutt R, Bonner–Weir S, and Weir GC. Chronic hyperglycemia triggers loss of pancreatic beta cell differentiation in an animal model of diabetes. J Biol Chem 274: 14112–14121, 1999.
- Jordan W, Cohrs S, Degner D, Meier A, Rodenbeck A, Mayer G, Pilz J, Ruther E, Kornhuber J, and Bleich S. Evaluation of oxidative stress measurements in obstructive sleep apnea syndrome. J Neural Transm 113: 239–254, 2006.
- Joslin E. The prevention of diabetes mellitus. JAMA 76: 79–84, 1921.
- Kagan VE, Shvedova AA, Novikov KN, and Kozlov YP. Lightinduced free radical oxidation of membrane lipids in photoreceptors of frog retina. *Biochim Biophys Acta* 330: 76–79, 1973.
- 133. Kakimoto M, Inoguchi T, Sonta T, Yu HY, Imamura M, Etoh T, Hashimoto T, and Nawata H. Accumulation of 8-hydroxy-2'deoxyguanosine and mitochondrial DNA deletion in kidney of diabetic rats. *Diabetes* 51: 1588–1595, 2002.
- 134. Kanauchi M, Nishioka H, and Hashimoto T. Oxidative DNA damage and tubulointerstitial injury in diabetic nephropathy. *Nephron* 91: 327–329, 2002.
- Kaneto H, Xu G, Fujii N, Kim S, Bonner–Weir S, and Weir GC. Involvement of c-Jun N-terminal kinase in oxidative stress-mediated suppression of insulin gene expression. *J Biol Chem* 277: 30010–30018, 2002.
- 136. Kaneto H, Xu G, Song KH, Suzuma K, Bonner–Weir S, Sharma A, and Weir GC. Activation of the hexosamine pathway leads to deterioration of pancreatic beta-cell function through the induction of oxidative stress. *J Biol Chem* 276: 31099–31104, 2001.
- 137. Kaneto HH, Xu G, Fujii N, Kim S, Bonner-Weir S, and Weir GC. Involvement of protein kinase C beta 2 in c-myc induction by high glucose in pancreatic beta-cells. *J Biol Chem* 277: 3680–3685, 2002.
- Kannel WB and McGee DL. Diabetes and cardiovascular diseases: the Framingham Study. JAMA 241: 2035–2038, 1979.
- 139. Karowicz–Bilinska A, Suzin J, and Sieroszewski P. Evaluation of oxidative stress indices during treatment in pregnant women with intrauterine growth retardation. *Med Sci Monit* 8: CR211–CR216, 2002.
- 140. Keaney JF Jr, Larson MG, Vasan RS, Wilson PW, Lipinska I, Corey D, Massaro JM, Sutherland P, Vita JA, Benjamin EJ, Framingham Study. Obesity and systemic oxidative stress: clinical correlates of oxidative stress in the Framingham Study. Arterioscler Thromb Vasc Biol 23: 434–439, 2003.
- 141. Kerr S, Brosnan MJ, Mcintyre M, Reid JL, Dominiczak AF, and Hamilton CA. Superoxide anion production is increased in a model of genetic hypertension: role of the endothelium. *Hypertension* 33: 1353–1358, 1999.
- 142. Kikuchi H, Furuta A, Nishioka K, Suzuki SO, Nakabeppu Y, and Iwaki T. Impairment of mitochondrial DNA repair enzymes against accumulation of 8-oxo-guanine in the spinal motor neurons of amyotrophic lateral sclerosis. *Acta Neuropathol (Berl)* 103: 408–414, 2002.
- Kirpichnikov D, McFarlane SI, and Sowers JR. Metformin: an update. Ann Intern Med 137: 25–33, 2002.

- 144. Klebanoff. Peroxidases in Chemistry and Biology, edited by Everse, Everse, and Grisham. Boston, CRC Press. 1991, pp. 1–35.
- 145. Klein R, Klein BE, Moss SE, Davis MD, and DeMets DL. The Wisconsin epidemiologic study of diabetic retinopathy. III. Prevalence and risk of diabetic retinopathy when age at diagnosis is 30 or more years. Arch Ophthalmol 102: 27–32, 1984.
- 146. Klein R and Klein BE. Diabetic eye disease. Lancet 350: 197–204, 1997.
- 147. Knerr I, Wolf J, Reinehr T, Stachow R, Grabert M, Schober E, Rascher W, Holl RW; DPV Scientific Initiative of Germany and Austria. The 'accelerator hypothesis': relationship between weight, height, body mass index and age at diagnosis in a large cohort of 9,248 German and Austrian children with type 1 diabetes mellitus. *Diabetologia* 48: 2501–2504, 2005.
- Knight JA. Free radicals: their history and current status in aging and disease. Ann Clin Lab Sci 28: 331–346, 1998.
- Knight JA. Reactive oxygen species and the neurodegenerative disorders. Ann Clin Lab Sci 27: 11–25, 1997.
- Kosaka K, Noda M, and Kuzuya T. Prevention of type 2 diabetes by lifestyle intervention: a Japanese trial in IGT males. *Diabetes Res Clin Pract* 67: 152–162, 2005.
- Kowluru RA, Kern TS, Engerman RL, and Armstrong D. Abnormalities of retinal metabolism in diabetes or experimental galactosemia: III. Effects of antioxidants. *Diabetes* 45: 1233–1237, 1996
- 152. Kowluru RA, Kowluru V, Xiong Y, and Ho YS. Overexpression of mitochondrial superoxide dismutase in mice protects the retina from diabetes-induced oxidative stress. *Free Radic Biol Med* 41: 1191–1196, 2006.
- 153. Kowluru RA and Odenbach S. Effect of long-term administration of alpha lipoic acid on retinal capillary cell death and the development of retinopathy in diabetic rats. *Diabetes* 53: 3233–3238, 2004.
- 154. Kumar KV and Das UN. Are free radicals involved in the pathobiology of human essential hypertension? Free Radic Res. Commun 19: 59–66, 1993.
- 155. Kuroki M, Voest EE, Amano S, Beerepoot LV, Takashima S, Tolentino M, Kim RY, Rohan RM, Colby KA, Yeo KT, and Adamis AP. Reactive oxygen intermediates increase vascular endothelial growth factor expression in vitro and in vivo. *J Clin Invest* 98: 1667–1675, 1996.
- Laakso M. Hyperglycemia and cardiovascular disease in type 2 diabetes. *Diabetes* 48: 937–942, 1999.
- 157. Lacy F, Kailasam MT, O'Connor DT, Schmid–Schonbein GW, and Parmer RJ. Plasma hydrogen peroxide production in human essential hypertension: role of heredity, gender and ethnicity. Hypertension 36: 878–884, 2000.
- 158. Lakka HM, Laaksonen DE, Lakka TA, Niskanen LK, Kumpusalo E, Tuomilehto J, and Salonen JT. The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *JAMA* 288: 2709–2716, 2002.
- Lee AY and Chung SS. Contributions of polyol pathway to oxidative stress in diabetic cataract. FASEB J 13: 3–30, 1999.
- Lee HS, Son SM, Kim YK, Hong KW, and Kim CD. NAD(P)H oxidase participates in the signaling events in high glucoseinduced proliferation of vascular smooth muscle cells. *Life Sci* 72: 2719–2730, 2003.
- Leiter L and Lewanczuk RZ. Of the renin-angiotensin system and reactive oxygen species. Am J Hypertens 18: 121–128, 2005.
- Lenzen S, Drinkgern J, and Tiedge M. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. *Free Radic Biol Med* 20: 463–466, 1996.
- Leonard SS, Harris GK, and Shi XL. Metal-induced oxidative stress and signal transduction. Free Rad Biol Med 37: 1921–1942, 2004.
- 164. Liguori A, Abete P, Hayden JM, Cacciatore F, Rengo F, Ambrosio G, Bonaduce D, Condorelli M, Reaven PD, and Napoli C. Effect of glycaemic control and age on low-density lipoprotein susceptibility to oxidation in diabetes mellitus type 1. Eur Heart J 22: 2075–2084, 2001.
- Lin KY, Ito A, Asagami T, Tsao PS, Adimoolam S, Kimoto M, Tsuji H, Reaven GM, and Cooke JP. Impaired nitric oxide

- synthase pathway in diabetes mellitus: role of asymmetric dimethylarginine and dimethylarginine dimethylaminohydrolase. *Circulation* 106: 987–992, 2002.
- 166. Liochev SI and Fridovich I. The role of O2^{-*} in the production of HO*: in vitro and in vivo. Free Radic Biol Med 16: 29–33, 1994.
- Lou MF, Dickerson JE Jr, Garadi R, and York BM Jr. Glutathione depletion in the lens of galactosemic and diabetic rats. *Exp Eye Res* 46: 517–530, 1988.
- 168. Lu M, Kuroki M, Amano S, Tolentino M, Keough K, Kim I, Bucala R, and Adamis AP. Advanced glycation end products increase retinal vascular endothelial growth factor expression. *J Clin Invest* 101: 1219–1224, 1998.
- Lyras L, Cairns NJ, Jenner A, Jenner P, and Halliwell B. An assessment of oxidative damage to proteins, lipids, and DNA in brain from patients with Alzheimer's disease. *J Neurochem* 68: 2061–2069, 1997.
- 170. MacDonald-Wicks L, Gibson LZ, Godfrey DM, Green JM, Horan BP, Monger KL, Wischer RM, and Garg ML. Oxidised LDL in newly diagnosed type 2 diabetes mellitus and impaired glucose tolerance. Asia Pac J Clin Nutr 13(Suppl.): S65, 2004.
- 171. Maechler P, Jornot L, and Wollheim CB. Hydrogen peroxide alters mitochondrial activation and insulin secretion in pancreatic beta cells. *J Biol Chem* 274: 27905–27913, 1999.
- 172. Maedler K, Sergeev P, Ris F, Oberholzer J, Joller–Jemelka HI, Spinas GA, Kaiser N, Halban PA, and Donath MY. Glucose-induced β cell production of IL-1β contributes to glucotoxicity in human pancreatic islets. *J Clin Invest* 110: 851–860, 2002.
- 173. Maiorana A, O'Driscoll G, Cheetham C, Dembo L, Stanton K, Goodman C, Taylor R, and Green D. The effect of combined aerobic and resistance exercise training on vascular function in type 2 diabetes. *J Am Coll Cardiol* 38: 860–866, 2001.
- Makino A, Skelton MM, Zou AP. Roman RJ. and Cowley AW Jr. Increased renal medullary oxidative stress produces hypertension. *Hypertension* 39: 667–672, 2002.
- 175. Malaise WJ. Alloxan toxicity of pancreatic B-cell—a new hypothesis. *Biochem Pharmacol* 22: 3527–3534, 1982.
- 176. Malaisse WJ, Hutton JC, Carpinelli AR, Herchuelz A, and Senner A. The stimulus-secretion coupling of amino acidinduced insulin release. Metabolism and cationic effects of leucine. *Diabetes* 29: 431–437, 1980.
- 177. Malmberg K, Yusuf S, Gerstein H, Brown J, Zhao F, Hunt D, Piegas L, Calvin J, Keltai M, and Budaj A, for the OASIS Registry Investigators. Impact of diabetes on long-term prognosis in patients with unstable angina and non-Q-wave myocardial infarction: results of the OASIS (Organization to Assess Strategies for Ischemic Syndromes) Registry. Circulation 102: 1014–1019, 2000
- 178. Marfella R, Verrazzo G, Acampora R, La Marca C, Giunta R, Lucarelli C, Paolisso G, Ceriello A, and Giugliano D. Glutathione reverses systemic hemodynamic changes by acute hyperglycemia in healthy subjects. *Am J Physiol* 268: E1167–E1173, 1995.
- Martens FM, Visseren FL, Lemay J, de Koning EJ, and Rabelink TJ. Metabolic and additional effects of thiazolidinediones. *Drugs* 62: 1463–1480, 2002.
- 180. Mason RP, Walter MF, and Jacob RF. Effects of HMG-CoA reductase inhibitors on endothelial function: Role of microdomains and oxidative stress. *Circulation* 109: II-34–II-41, 2004
- Massy ZA and Nguyen–Khoa T. Oxidative stress and chronic renal failure: markers and management (Review). J Nephrol 15: 336–341, 2002.
- 182. McFarlane SI, Jacober SJ, Winer N, Kaur J, Castro JP, Wul MA, Gliwa A, Von Gizycki H, and Sowers JR. Control of cardiovascular risk factors in patients with diabetes and hypertension at urban academic medical centers. *Diabetes Care* 25: 718–723, 2002
- 183. McFarlane SI, Muniyappa R, Francisco R, and Sowers JR. Clinical review 145: Pleiotropic effects of statins: lipid reduction and beyond. *J Clin Endocrinol Metab* 87: 1451-1458, 2002.
- 184. McFarlane SI, Shin JJ, Rundek T, and Bigger JT. Prevention of type 2 diabetes. *Curr Diab Rep* 3: 235–241, 2003.
- Mehta JL, Hu B, Chen J, and Li D. Pioglitazone inhibits LOX-1 expression in human coronary artery endothelial cells by reduc-

- ing intracellular superoxide radical generation. Arterioscl Thromb Vasc Biol 23: 2203–2208, 2003.
- 186. Mehta JL, Rasouli N, Sinha AK, and Molavi B. Oxidative stress in diabetes: A mechanistic overview of its effects on atherogenesis and myocardial dysfunction. *Int J Biochem Cell Biol* 38: 794–803, 2006.
- 187. Mendola J, Wright Jr JR, and Lacy PE. Oxygen free radical scavengers and immune destruction of murine islets in allograft rejection and multiple low dose streptozocin induced insulitis. *Diabetes* 38: 379–385, 1989.
- 188. Mertz RJ, Worley JF-III, Spencer B, Johnson JH, and Dukes ID. Activation of stimulus-secretion coupling in pancreatic β-cells by specific products of glucose metabolism. *J Biol Chem* 271: 4838–4845, 1996.
- 189. Metodiewa D and Koska C. Reactive oxygen species and reactive nitrogen species: relevance to cyto(neuro)toxic events and neurologic disorders. An overview. *Neurotox Res* 1: 197–233, 2000.
- Miettinen HL, Salomaa S, Mahonen V, Niemela M, Haffner M, Pyorala M, and Tuomilehto K, for the FINMONICA Myocardial Infarction Register Study Group. Impact of diabetes on mortality after the first myocardial infarction. *Diabetes Care* 21: 69–75, 1998.
- 191. Miller JW, Adamis AP, Shima DT, D'Amore PA, Moulton RS, O'Reilly MS, Folkman J, Dvorak HF, Brown LF, Berse B, Yeo T, and Yeo K. Vascular endothelial growth factor/vascular permeability factor is temporally and spatially correlated with ocular angiogenesis in a primate model. *Am J Pathol* 145: 574–584, 1994.
- Mohan IK and Das UN. Oxidant stress, anti-oxidants and essential fatty acids in systemic lupus erythematosus. *Prostaglandins Leukot Essent Fatty Acids* 56: 193–198, 1997.
- 193. Monnier VM, Bautista O, Kenny D, Sell DR, Fogarty J, Dahms W, Cleary PA, Lachin J, and Genuth S. Skin collagen glycation, glycoxidation, and crosslinking are lower in subjects with long-term intensive versus conventional therapy of type 1 diabetes: relevance of glycated collagen products versus HbA1c as markers of diabetic complications. DCCT Skin Collagen Ancillary Study Group. Diabetes Control and Complications Trial. *Diabetes* 48: 870–880, 1999.
- 194. Moran A, Zhang HJ, Olsonm LK, Harmon JS, Poitoust V, and Robertson RP. Differentiation of glucose toxicity from β-cell exhaustion during the evolution of defective insulin gene expression in the pancreatic islet cell line, HIT-T15. *J Clin Invest* 99: 534–539, 2000.
- 195. Morrow JD, Awad JA, Kato T, Takahashi K, Badr KF, Roberts LJ 2nd, and Burk RF. Formation of novel non-cyclooxygenasederived prostanoids (F2-isoprostanes) in carbon tetrachloride hepatotoxicity. An animal model of lipid peroxidation. *J Clin Invest* 90: 2502–2507, 1992.
- 196. Moustafa SA. Zinc might protect oxidative changes in the retina and pancreas at the early stage of diabetic rats. *Toxicol Appl Pharmacol* 201: 149–155, 2004.
- Nakazono K, Watanabe N, Matsuno K, Sasaki J, SatoT, and Inoue M. Does superoxide underlie the pathogenesis of hypertension? *Proc Natl Acad Sci USA* 88: 10045–10048, 1991.
- Nathan DM. Long-term complications of diabetes mellitus. N Engl J Med 328: 1676–1685, 1993.
- Negita M, Yokoyama I, Hayashi S, Kobayashi T, Yasutomi M, and Takagi H. Superoxide scavenging activity in experimental liver transplantation. *Transpl Int* 8: 256–261, 1995.
- Newgard CB and McGarry JD. Metabolic coupling factors in pancreatic β-cell signal transduction. Ann Rev Biochem 64: 689–719, 1995.
- Nicasio J and McFarlane SI. Early insulin therapy and the risk of cardiovascular disease in type 2 diabetes. *Therapy* 5: 685–688, 2005
- 202. Ninomiya JK, L'Italien G, Criqui MH, Whyte JL, Gamst A, and Chen RS. Association of the metabolic syndrome with history of myocardial infarction and stroke in the Third National Health and Nutrition Examination Survey. *Circulation* 109: 42–46, 2004.
- Nishikawa T, Edelstein D, Du X-L, Yamagishi S, Matsumura T, Kaneda Y, Yorek M, Beebe D, Oates P, Hammes HP, Giardino I,

- and Brownlee M. Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* 404: 787–790, 2000.
- 204. Noda M, Yamashita S, Takahashi N, Eto K, Shen LM, Izumi K, Daniel S, Tsubamoto Y, Nemoto T, Lino M, Kasai H, Sharp GW, and Kadowaki T. Switch to anaerobic glucose metabolism with NADH accumulation in the beta-cell model of mitochondrial diabetes. Characteristics of betaHC9 cells deficient in mitochondrial DNA transcription. J Biol Chem 277: 41817–41826, 2002.
- Noda Y, Mori A, and Packer L. Gliclazide scavengers hydroxyl, superoxide and nitric oxide radicals; An ESR study. Res Commun Mol Pathol Pharmacol 96: 115–124, 1997.
- O'Donnell RW, Johnson DK, Ziegler LM, DiMattina AJ, Stone RI, and Holland JA. Endothelial NADPH oxidase: mechanism of activation by low-density lipoprotein. *J Endothel Cell Res* 10: 291–297, 2003.
- Oberley LW. Free radicals and diabetes. Free Rad Bio Med 5: 113–124, 1988.
- 208. Obrosova IG, Minchenko AG, Vasupuram R, White L, Abatan OI, Kumagai AK, Frank RN, and Stevens MJ. Aldose reductase inhibitor fidarestat prevents retinal oxidative stress and vascular endothelial growth factor overexpression in streptozotocindiabetic rats. *Diabetes* 52: 864–871, 2003.
- Obrosova IG and Julius UA. Role for poly (ADP-ribose) polymerase activation in diabetic nephropathy, neuropathy and retinopathy. Curr Vasc Pharmacol 3: 267–283, 2005.
- Odetti P, Traverso N, Cosso L, Noberasco G, Pronzato MA, and Marinari UM. Good glycaemic control reduces oxidation and glycation end-products in collagen of diabetic rats. *Diabetologia* 39: 1440–1447, 1996.
- 211. Olusi SO. Obesity is an independent risk factor for plasma lipid peroxidation and depletion of erythrocyte cytoprotective enzymes in humans. *Int J Obes Relat Metab Disord* 26: 1159–1164, 2002.
- Omar BA and McCord JM. Interstitial equilibration of superoxide dismutase correlates with its protective effect in the isolated rabbit heart. *J Mol Cell Cardiol* 23: 149–159, 1991.
- 213. Ozawa T, Hayakawa M, Katsumata K, Yoneda M, Ikebe S, and Mizuno Y. Fragile mitochondrial DNA: the missing link in the apoptotic neuronal cell death in Parkinson's disease. *Biochem Biophys Res Commun* 235: 158–161, 1997.
- 214. Pan XR, Li GW, Hu YH, Wang JX, Yang WY, An ZX, Hu ZX, Lin J, Xiao JZ, Cao HB, Liu PA, Jiang XG, Jiang YY, Wang JP, Zheng H, Zhang H, Bennett PH, and Howard BV. Effects of diet and exercise in preventing NIDDM in people with impaired glucose tolerance: the Da Qing IGT and Diabetes Study. *Diabetes Care* 20: 537–544, 1997.
- Panten U, Zielman S, Langer J, Zunkler BJ, and Lenzen S. Regulation of insulin secretion by energy metabolism in pancreatic β-cell mitochondria. *Biochem J* 219: 189–196, 1984.
- Parks DA, Bulkley GB, Granger DN, Hamilton SR, and McCord JM. Ischemic injury in the cat small intestine: role of superoxide radicals. *Gastroenterology* 82: 9–15, 1982.
- 217. Parthasarathy S, Fong LC, and Steinberg D. Oxidative modification of low density lipoprotein and atherosclerosis: concepts and consequences. In: *Lipid Peroxidation in Biological Systems*, edited by Sevanian A. Champaign, IL. Am Oil Chem Soc 1987, pp. 225–235.
- 218. Pastor N, Weinstein H, Jamison E, and Brenowitz M. A detailed interpretation of OH radical footprints in a TBP–DNA complex reveals the role of dynamics in the mechanism of sequencespecific binding. *J Mol Biol* 304: 55–68, 2000.
- Pedro–Botet J, Covas MI, Martin S, and Rubies–Prat J. Decreased endogenous antioxidant enzymatic status in essential hypertension. J Hum Hypertens 14: 343–345, 2000.
- 220. Peiro C, Lafuente N, Matesanz N, Cercas E, Llergo JL, Vallejo S, Rodriguez–Manas L, and Sanchez–Ferrer CF. High glucose induces cell death of cultured human aortic smooth muscle cells through the formation of hydrogen peroxide. *Br J Pharmacol* 133: 967–974, 2001.
- Perkins ND. The Rel/NF-κB family: friend and foe. *Trends Biochem Sci* 25: 434–440, 2000.

- Petersen KF, Dufour S, Befroy D, Garcia R, and Shulman GI. Impaired mitochondrial activity in the insulin-resistant offspring of patients with type 2 diabetes. N Engl J Med 350: 664–667, 2004
- 223. Peuchant E, Delmas—Beauvieux MC, Couchouron A, Dubourg L, Thomas MJ, Perromat A, Clerc M, and Gin H. Short -term insulin therapy and normoglycemia: effects on erythrocyte lipid peroxidation in NIDDM patients. *Diabetes Care* 20: 202–207, 1997.
- Pieper GM and Siebeneich W. Oral administration of the antioxidant, N-acetylcysteine, abrogates diabetes-induced endothelial dysfunction. *J Cardiovasc Pharmacol* 32: 101–105, 1998.
- Polydefkis M, Griffin JW, and McArthur J. New insights into diabetic polyneuropathy. *JAMA* 290: 1371–1376, 2003.
- Pryor WA and Church DF. Aldehydes, hydrogen peroxide and organic radicals as mediators of ozone toxicity. Free Rad Biol Med 11: 41–46, 1991.
- Ragusa RJ, Chow CK, and Porter JD. Oxidative stress as a potential pathogenic mechanism in an animal model of Duchenne muscular dystrophy. *Neuromuscul Disord* 7: 379–386, 1997.
- Rahimi R, Nikfar S, Larijani B, and Abdollahi M. A review on the role of antioxidants in the management of diabetes and its complications. *Biomed Pharmacother* 59: 365–373, 2005.
- Reaven GM. Banting lecture 1988. Role of insulin resistance in human disease. *Diabetes* 37: 1595–1607, 1988.
- Redon J, Oliva MR, Tormos C, Giner V, Chaves J, Iradi A, and Saez GT. Antioxidant activities and oxidative stress byproducts in human hypertension. *Hypertension* 41: 1096–1101, 2003.
- Ricote M, Li A–C, Wilson T–M, Kelly C–J, and Glass C–K. The peroxisome proliferator-activated receptor-γ is a negative regulator of macrophage activation. *Nature* 391: 79–82, 1998.
- Steinbrook R. Facing the diabetes epidemic—mandatory reporting of glycosylated hemoglobin values in New York City. N Engl J Med 354: 545–548, 2006.
- Roberts CK and Barnard RJ. Effects of exercise and diet on chronic disease. J Appl Physiol 98: 3–30, 2005.
- Roberts CK, Massey MR, Cherny S, O'Brien LT, and Pritikin N. Long-term use of a high-complex-carbohydrate, high-fiber, low fat diet and exercise in the treatment of NIDDM patients. *Dia*betes Care 6: 268–273, 1983.
- 235. Roberts CK, Won D, Pruthi S, Lin SS, and Barnard RJ. Effect of a diet and exercise intervention on oxidative stress, inflammation and monocyte adhesion in diabetic men. *Diabetes Res Clin Pract* 73: 249–259, 2006.
- Robertson RP. Chronic oxidative stress as a central mechanism for glucose toxicity in pancreatic islet beta cells in diabetes. *J Biol Chem* 41: 42351–42354, 2004.
- Roglic G, Unwin N, Bennett PH, Mathers C, Tuomilehto J, Nag S, Connolly V, and King H. The burden of mortality attributable to diabetes: realistic estimates for the year 2000. *Diabetes Care* 28: 2130–5, 2005.
- Rosca MG, Monnier VM, Szweda LI, and Weiss MF. Alterations in renal mitochondrial respiration in response to the reactive oxoaldehyde methylglyoxal. *Am J Physiol Renal Physiol* 283: F52–F59, 2002.
- 239. Rosen P, Nawroth PP, King G, Moller W, Tritschler HJ, and Packer L. The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes Metab* Res Rev 17: 189–212, 2001.
- Ross R. Atherosclerosis—an inflammatory disease. N Eng J Med 340: 115–126, 2004.
- Rubler S, Dlugash J, Yuceoglu YZ, Kumral T, Branwood AW, and Grishman A. New type of cardiomyopathy associated with diabetic glomerulosclerosis. Am J Cardiol 30: 595–602, 1972.
- Russell JW, Golovoy D, Vincent AM, Mahendru P, Olzmann JA, Mentzer A, and Feldman EL. High glucose-induced oxidative stress and mitochondrial dysfunction in neurons. *FASEB J* 16: 1738–1748, 2002.
- Russell JW, Sullivan KA, Windebank AJ, Herrmann DN, and Feldman EL. Neurons undergo apoptosis in animal and cell culture models of diabetes. *Neurobiol Dis* 6: 347–363, 1999.

- 244. Sakai K, Matsumoto K, Nishikawa T, Suefuji M, Nakamaru K, Hirashima Y, Kawashima J, Shirotani T, Ichinose I, Brownlee M, and Araki E. Mitochondrial reactive oxygen species reduce insulin secretion by pancreatic β-cells. *Biochem Biophys Res* Commun 300: 216–222, 2003.
- 245. Sampaio RC, Tanus–Santos JE, Melo SE, Hyslop S, Franchini KG, Luca IM, and Moreno H Jr. Hypertension plus diabetes mimics the cardiomyopathy induced by nitric oxide inhibition in rats. *Chest* 122: 1412–1420, 2002.
- 246. Santini SA, Marra G, Giardina B, Cotroneo P, Mordente A, Martorana GE, Manto A, and Ghirlanda G. Defective plasma antioxidant defenses and enhanced susceptibility to lipid peroxidation in uncomplicated IDDM. *Diabetes* 46: 1853–1858, 1997.
- Sastre J, Pallardo FV, Pla R, Pellin A, Juan G, O'Connor JE, Estrela JM, Miquel J, and Vina J. Aging of the liver: age-associated mitochondrial damage in intact hepatocytes. *Hepatology* 24: 1199–1205, 1996.
- Sato Y, Hotta N, Sakamoto N, Matsuoka S, Ohishi N, and Yagi K. Lipid peroxide level in plasma of diabetic patients. *Biochem Med* 21: 104–107, 1979.
- Saydah SH, Fradkin J, and Cowie CC. Poor control of risk factors for vascular disease among adults with previously diagnosed diabetes. *JAMA* 291: 335–342, 2004.
- Schleicher ED, Wagner E, and Nerlich AG. Increased accumulation of the glycoxidation product Ne-(carboxymethyl) lysine in human tissues in diabetes and aging. *J Clin Invest* 99: 457–468, 1997.
- Schmeichel AM, Schmelzer JD, and Low PA. Oxidative injury and apoptosis of dorsal root ganglion neurons in chronic experimental diabetic neuropathy. *Diabetes* 52: 165–171, 2003.
- 252. Schuit F. Metabolic fate of glucose in purified islet cells. Glucose regulated anaplerosis in β-cells. *J Biol Chem* 272: 18572–18579, 1997.
- Selak MA, Storey BT, Peterside IE, and Simmons RA. Impaired oxidative phosphorylation in skeletal muscle contributes to insulin resistance and hyperglycemia. Am J Physiol 285: E130–E137, 2003.
- Shah SV. The role of reactive oxygen metabolites in glomerular disease. *Annu Rev Physiol* 57: 245–262, 1995.
- Shepherd PR and Kahn BB. Glucose transporters and insulin action. Implications for insulin resistance and diabetes mellitus. N Eng J Med 341: 248–257, 1999.
- Sies H, ed. Oxidative Stress II: Oxidants and Antioxidants. New York: Academic Press, 1991.
- Simmons RA. Developmental origins of diabetes: The role of oxidative stress. Free Radical Biol Med 40: 917–922, 2006.
- 258. Simoneau JA, Colberg SR, Thaete FL, and Kelley DE. Skeletal muscle glycolytic and oxidative enzyme capacities are determinants of insulin sensitivity and muscle composition in obese women. FASEB J 9: 273–278, 1995.
- Skyrme–Jones RA. O'Brien RC, Berry KL, and Meredith IT. Vitamin E supplementation improves endothelial function in type I diabetes mellitus: a randomized, placebo-controlled study. *J Am Coll Cardiol* 36: 94–102, 2000.
- 260. Spiekermann S, Landmesser U, Dikalov S, Bredt M, Gamez G, Tatge H, Reepschlager N, Hornig B, Drexler H, and Harrison DG. Electron spin resonance characterization of vascular xanthine and NAD(P)H oxidase activity in patients with coronary artery disease. *Circulation* 107: 1383, 2003.
- Stadtman ER and Oliver DN. Metal-catalyzed oxidation of proteins. Physiological consequences. *J Biol Chem* 266: 2005–2008, 1001
- 262. Steinberg D, Parthasarathy S, Carew TE, Khoo JC, and Witztum JL. Beyond cholesterol: modifications of low density lipoproteins that increase its atherogenicity. N Engl J Med 320: 915–924, 1989.
- 263. Stocks J, Gutteridge JM, Sharp RJ, and Dormandy TL. The inhibition of lipid autoxidation by human serum and its relation to serum proteins and α-tocopherol. Clin Sci Mol Med 47: 223–233, 1974.
- 264. Stumvoll M and Haring H–U. Glitazones: clinical effects and molecular mechanisms. *Ann Med* 34: 217–222, 2002.

265. Sugimoto H, Shikata K, Wada J, Horiuchi S, and Makino H. Advanced glycation end products-cytokine-nitric oxide sequence pathway in the development of diabetic nephropathy: aminoguanidine ameliorates the overexpression of tumour necrosis factor-alpha and inducible nitric oxide synthase in diabetic rat glomeruli. *Diabetologia* 42: 878–886, 1999.

- 266. Suzuki D, Miyata T, Saotome N, Horie K, Inagi R, Yasuda Y, Uchida K, Izuhara Y, Yagama M, Sakai H, and Kurokawa K. Immunohistochemical evidence for an increased oxidative stress and carbonyl modification of proteins in diabetic glomerular lesions. *J Am Soc Nephrol* 10: 822–832, 1999.
- Suzuki LA, Poot M, Gerrity RG, and Bornfeldt KE. Diabetes accelerates smooth muscle accumulation in lesions of atherosclerosis: lack of direct growth-promoting effects of high glucose levels. *Diabetes* 50: 851–860, 2001.
- 268. Suzuki T, Tatsuoka H, Chiba T, Sekikawa T, Nemoto T, Moriya H, Sakuraba S, and Nakaya H. Beneficial effects of nitric oxide synthase inhibition on the recovery of neurological function after spinal cord injury in rats. *Naunyn Schmiedebergs Arch Pharmacol* 363: 94–100, 2001.
- Takaishi H, Taniguchi T, Takahashi A, Ishikawa Y, and Yokoyama M. High glucose accelerates MCP-1 production via p38 MAPK in vascular endothelial cells. *Biochem Biophys Res Commun* 305: 122–128, 2003.
- Tanaka Y, Tran PO, Harmon J, and Robertson RP. A role for glutathione peroxidase in protecting pancreatic βcells against oxidative stress in a model of glucose toxicity. *Proc Natl Acad Sci USA* 99: 12363–12368, 2002.
- Tesfamariam B and Cohen RA. Free radicals mediate endothelial cell dysfunction caused by elevated glucose. *Am J Physiol* 263: H321–H326, 1992.
- 272. The Diabetes Control and Complications Trials (DCCT) Research Group. The effect of intensive treatment of diabetes on the development and progression of longterm complications in insulin-dependent diabetes mellitus. N Engl J Med 329: 977–986, 1993.
- The DREAM Trial Investigators. Effect of ramipril on the incidence of diabetes. N Engl J Med 355: 1551–1562, 2006.
- 274. Thompson HJ, Heimendinger J, Haegele A, Sedlacek SM, Gillette C, O'Neill C, Wolfe P, and Conry C. Effect of increased vegetable and fruit consumption on markers of oxidative cellular damage. *Carcinogenesis* 20: 2261–2266, 1999.
- 275. Tiedge M, Lortz S, Munday R, and Lenzen S. Protection against the cooperative toxicity of nitric oxide and oxygen free radicals by overexpression of antioxidant enzymes in bioengineered insulin-producing RINm5F cells. *Diabetologia* 42: 849–855, 1999
- Tiedge M, Lortz S, Drinkgern J, and Lenzen S. Relationship between antioxidant enzyme gene expression and antioxidant defense status of insulin-producing cells. *Diabetes* 46: 1733–1742, 1997.
- 277. Tin RG, Chang K, Nyengaard JR, van den Enden M, Ido Y, and Williamson JR. Inhibition of sorbitol dehydrogenase. Effects on vascular and neural dysfunction in streptozocin-induced diabetic rats. *Diabetes* 44: 234–242, 1995.
- Ting HH, Timimi FK, Boles KS, Creager SJ, Ganz P, and Creager MA. Vitamin C improves endothelium-dependent vasodilation in patients with non-insulin-dependent diabetes mellitus. *J Clin Invest* 97: 22–28, 1996.
- Toshniwal PK and Zarling EJ. Evidence for increased lipid peroxidation in multiple sclerosis. *Neurochem Res* 17: 205–207, 1992.
- 280. Toyokuni S, Tanaka T, Hattori Y, Nishiyama Y, Yoshida A, Uchida K, Hiai H, Ochi H, and Osawa T. Quantitative immuno-histochemical determination of 8-hydroxy-2'-deoxyguanosine by a monoclonal antibody N45.1: its application to ferric nitrilotriacetate-induced renal carcinogenesis model. *Lab Invest* 76: 365–374, 1997.
- Trachtman H, Futterweit S, Pine E, Mann J, and Valderama E. Chronic diabetic nephropathy: role of inducible nitric oxide synthase. *Pediatr Nephrol* 17: 20–29, 2002.
- Tuomilehto J, Lindstrom J, Eriksson JG, Valle TT, Hamalainen H, Ilanne-Parikka P, Keinanen-Kiukaanniemi S, Laakso M,

- Louheranta A, Rastas M, Salminen V, and Uusitupa M, the Finnish Diabetes Prevention Study Group. Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med* 344: 1343–1350, 2001.
- 283. UK Prospective Diabetes Study Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* 352: 837–853, 1998.
- Urso ML and Clarkson PM. Oxidative stress, exercise, and antioxidant supplementation. *Toxicology* 189: 41–54, 2003.
- Vaille A, Jadot G, and Elizagaray A. Anti-inflammatory activity
 of various superoxide dismutases on polyarthritis in the Lewis
 rat. *Biochem Pharmacol* 39: 247–255, 1990.
- Valko M, Rhodes CJ, Moncola J, Izakovic M, and Mazur M. Free radicals, metals & antioxidants in oxidative stress-induced cancer. *Chemico-Biol Interactions* 160: 1–40, 2006.
- 287. Van den Enden MK, Nyengaard JR, Ostrow E, Burgan JH, and Williamson JR. Elevated glucose levels increase retinal glycolysis and sorbitol pathway metabolism: implications for diabetic retinopathy. *Invest Ophthalmol Vis Sci* 36: 1675–1685, 1995.
- Vecchione C and Brandes RP. Withdrawal of the 3-hydroxy-3-methylglutaryl coenzyme A reductase inhibitors elicits oxidative stress and induces endothelial dysfunction in mice. *Circ Res* 91: 173–179, 2002.
- Venugopal SK, Devaraj S, Yang T, and Jialal I. Alpha-tocopherol decreases superoxide anion release in human monocytes under hyperglycemic conditions via inhibition of protein kinase C-alpha. *Diabetes* 51: 3049–3054, 2002.
- 290. Verzola D, Bertolotto MB, Villaggio B, Ottonello L, Dallegri F, Frumento G, Berruti V, Gandolfo MT, Garibotto G, and Deferran G. Taurine prevents apoptosis induced by high ambient glucose in human tubule renal cells. *J Investig Med* 50: 443–451, 2002.
- Vincent AM, McLean LL, Backus C, and Feldman EL. Shortterm hyperglycemia produces oxidative damage and apoptosis in neurons. *FASEB J* 19: 638–640, 2005.
- 292. Vincent AM, Russell JW, Low P, and Feldman E. Oxidative stress in the pathogenesis of diabetic neuropathy. *Endocr Rev* 25: 612–628, 2004.
- 293. Vinik AI, et al. Diabetic autonomic neuropathy. In: *Ellenberg and Rifkin's Diabetes Mellitus*, edited by Porte D Jr, Sherwin RS, and Baron A. New York, McGraw Hill. 2002, pp. 789–804.
- Wallaert B, Aerts C, Gressier B, Gosset P, and Voisin C. Oxidative inactivation of alpha(1)-proteinase inhibitor by alveolar epithelial type II cells. *J Appl Physiol* 75: 2376–2382, 1993.
- 295. Warren JS, Yabroff KR, Mandel DM, Johnson KJ, and Ward PA. Role of O2 in neutrophil recruitment into sites of dermal and pulmonary vasculitis. Free Radical Biol Med 8: 163–172, 1990.
- 296. Wassmann S, Laufs U, Baumer AT, Muller K, Konkol C, Sauer H, Bohm M, and Nickenig G. Inhibition of geranylgeranylation reduces angiotensin II-mediated free radical production in vascular smooth muscle cells: involvement of angiotensin AT-1-receptor expression and rac1 GTPase. *Mol Pharmacol* 59: 646–654, 2001.
- Wautier JL, Wautier MP, Schmidt AM, Anderson GM, Hori O, Zoukourian C, Capron L, Chappey O, Tan SD, and Brett J.

- Advanced glycation end products (AGEs) on the surface of diabetic erythrocytes bind to the vessel wall via a specific receptor inducing oxidant stress in the vasculature: a link between surface- associated AGEs and diabetic complications. *Proc Natl Acad Sci USA* 91: 7742–7746, 1994.
- Wautier MP, Chappey O, Corda S, Stern DM, Schmidt AM, and Wautier JL. Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE. Am J Physiol Endocrinol Metab 280: E685–E694, 2001.
- Williams S, Cusco J, Roddy M, Johnstone M, and Creager M. Impaired nitric oxide-mediated vasodilation in patients with noninsulin-dependent diabetes mellitus. *J Am Coll Cardiol* 27: 567–574, 1996.
- Williamson JR, Chang K, Frangos M, Hasan KS, Ido Y, Kawamura T, Nyengaard JR, van den Enden M, Kilo C, and Tilton RG. Hyperglycemic pseudohypoxia and diabetic complications. *Diabetes* 42: 801–813, 1993.
- Wolff SP and Dean RT. Glucose autoxidation and protein modification. The potential role of 'autoxidative glycosylation' in diabetes. *Biochem J* 245: 243–250, 1987.
- Wolff SP. Diabetes mellitus and free radicals. Free radicals, transition metals and oxidative stress in the aetiology of diabetes mellitus and complications. Br Med Bull 49: 642–652, 1993
- Yim MB, Yim HS, Lee C, Kang SO, and Chock PB. Protein glycation: creation of catalytic sites for free radical generation. *Ann* NY Acad Sci 928: 48–53, 2001.
- 304. Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, and Dagenais G, the Heart Outcomes Prevention Evaluation Study Investigators. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. N Engl J Med 342: 145–153, 2000.
- Zalba G, San Jose G, Moreno MU, Fortuno MA, Fortuno A, Beaumont FJ, and Diez J. Oxidative stress in arterial hypertension. Role of NAD(P)H oxidase. *Hypertension* 38: 1395–1399, 2001.
- Zamocky M and Koller F. Understanding the structure and function of catalases: clues from molecular evolution and *in vitro* mutagenesis. *Prog Biophys Mol Biol* 72: 19–66, 1999.

Address reprint requests to:
Samy I. McFarlane, M.D., M.P.H., F.A.C.P.
Professor of Medicine
Chief, Division of Endocrinology, Diabetes and Hypertension
SUNY-Downstate Medical Center and Kings County
Hospital Center
450 Clarkson Avenue, Box 50
Brooklyn, New York 11203

E-mail: smcfarlane@downstate.edu

Date of first submission to ARS Central, February 26, 2007; date of acceptance, February 27, 2007.

This article has been cited by:

- 1. Christopher Papandreou, Sophia E. Schiza, Manolis N. Tzatzarakis, Mathaios Kavalakis, Christos M. Hatzis, Aristidis M. Tsatsakis, Anthony G. Kafatos, Nikolaos M. Siafakas, Nikolaos E. Tzanakis. 2012. Effect of Mediterranean diet on lipid peroxidation marker TBARS in obese patients with OSAHS under CPAP treatment: a randomised trial. *Sleep and Breathing* 16:3, 873-879. [CrossRef]
- 2. Pilar Codoñer-Franch, Almudena Navarro-Ruiz, María Fernández-Ferri, Ángela Arilla-Codoñer, Esther Ballester-Asensio, Victoria Valls-Bellés. 2012. A matter of fat: insulin resistance and oxidative stress. *Pediatric Diabetes* 13:5, 392-399. [CrossRef]
- 3. Guy Cohen, Yael Riahi, Shlomo SassonFree Radicals and Metabolic Disorders . [CrossRef]
- 4. Massimo D'Archivio, Giovanni Annuzzi, Rosaria Varì, Carmelina Filesi, Rosalba Giacco, Beatrice Scazzocchio, Carmela Santangelo, Claudio Giovannini, Angela A. Rivellese, Roberta Masella. 2011. Predominant role of obesity/insulin resistance in oxidative stress development. *European Journal of Clinical Investigation* no-no. [CrossRef]
- 5. Kenneth S. Hershon. 2011. Mechanistic and Clinical Aspects of Renin-Angiotensin-Aldosterone System Blockade in the Prevention of Diabetes Mellitus and Cardiovascular Disease. *Endocrine Practice* 17:3, 430-440. [CrossRef]
- 6. Samy I. McFarlane, Peter A. McCullough, James R. Sowers, Kyaw Soe, Shu-Cheng Chen, Suying Li, Joseph A. Vassalotti, Lesley A. Stevens, Moro O. Salifu, Manjula Kurella Tamura, Andrew S. Bomback, Keith C. Norris, Allan J. Collins, George L. Bakris, Adam T. Whaley-Connell. 2011. Comparison of the CKD Epidemiology Collaboration (CKD-EPI) and Modification of Diet in Renal Disease (MDRD) Study Equations: Prevalence of and Risk Factors for Diabetes Mellitus in CKD in the Kidney Early Evaluation Program (KEEP). American Journal of Kidney Diseases 57:3, S24-S31. [CrossRef]
- 7. Huimei Chen, Caixia Sun, Wenwen Guo, Ran Meng, Hong Du, Qiufeng Qi, Xin Gu, Lirong Li, Kui Zhang, Dalong Zhu. 2011. AluYb8 insertion in the MUTYH gene is related to increased 8-OHdG in genomic DNA and could be a risk factor for type 2 diabetes in a Chinese population. *Molecular and Cellular Endocrinology* 332:1-2, 301-305. [CrossRef]
- 8. Xiao-dong Zhang, Jian-wei Yan, Gui-rui Yan, Xiao-yun Sun, Jun Ji, Yi-ming Li, You-hong Hu, He-yao Wang. 2010. Pharmacological inhibition of diacylglycerol acyltransferase 1 reduces body weight gain, hyperlipidemia, and hepatic steatosis in db/db mice. *Acta Pharmacologica Sinica* 31:11, 1470-1477. [CrossRef]
- 9. Arlene A Stecenko, Antoinette Moran. 2010. Update on cystic fibrosis-related diabetes. *Current Opinion in Pulmonary Medicine* **16**:6, 611-615. [CrossRef]
- 10. Mohamed M. Elseweidy, Mona M. Taha, Nahla N. Younis, Khadiga S. Ibrahim, Hamdi A. Hamouda, Mohamed A. Eldosouky, Hala Soliman. 2010. Gastritis Induced by Helicobacter pylori Infection in Experimental Rats. *Digestive Diseases and Sciences* **55**:10, 2770-2777. [CrossRef]
- 11. Agnès Matheson, Mark D. P. Willcox, Judith Flanagan, Bradley J. Walsh. 2010. Urinary biomarkers involved in type 2 diabetes: a review. Diabetes/Metabolism Research and Reviews 26:3, 150-171. [CrossRef]
- 12. Itamar Yehuda, Zecharia Madar, Andrea Szuchman-Sapir, Snait Tamir. 2010. Glabridin, a phytoestrogen from licorice root, up-regulates manganese superoxide dismutase, catalase and paraoxonase 2 under glucose stress. *Phytotherapy Research* n/a-n/a. [CrossRef]
- 13. Olga P García, Kurt Z Long, Jorge L Rosado. 2009. Impact of micronutrient deficiencies on obesity. *Nutrition Reviews* **67**:10, 559-572. [CrossRef]
- 14. D. Reilly, C. A. Boyle, D. C. Craig. 2009. Obesity and dentistry: a growing problem. BDJ 207:4, 171-175. [CrossRef]
- 15. Noriko Yamabe , Ki Sung Kang , Jong Moon Hur , Takako Yokozawa . 2009. Matcha, a Powdered Green Tea, Ameliorates the Progression of Renal and Hepatic Damage in Type 2 Diabetic OLETF Rats. *Journal of Medicinal Food* 12:4, 714-721. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 16. Sushil K. Jain, Thirunavukkarasu Velusamy, Jennifer L. Croad, Justin L. Rains, Rebeca Bull. 2009. I-Cysteine supplementation lowers blood glucose, glycated hemoglobin, CRP, MCP-1, and oxidative stress and inhibits NF-#B activation in the livers of Zucker diabetic rats. *Free Radical Biology and Medicine* **46**:12, 1633-1638. [CrossRef]
- 17. Meriem Mahrouf-Yorgov, Nicolas Marie, Didier Borderie, Raja Djelidi, Dominique Bonnefont-Rousselot, Alain Legrand, Jean-Louis Beaudeux, Jacqueline Peynet. 2009. Metformin suppresses high glucose–induced poly(adenosine diphosphate–ribose) polymerase overactivation in aortic endothelial cells. *Metabolism* 58:4, 525-533. [CrossRef]
- 18. Andrew C. Miller, Brandon Smaglo, Samy I. McFarlane. 2009. Obesity, anemia, and cardiovascular risk in patients with chronic kidney disease: Overview and pathophysiologic insights. *Current Cardiovascular Risk Reports* **3**:2, 102-108. [CrossRef]
- 19. Sushil K. Jain, Justin Rains, Jennifer Croad, Bryon Larson, Kimberly Jones. 2009. Curcumin Supplementation Lowers TNF-#, IL-6, IL-8, and MCP-1 Secretion in High Glucose-Treated Cultured Monocytes and Blood Levels of TNF-#, IL-6, MCP-1, Glucose, and Glycosylated Hemoglobin in Diabetic Rats. *Antioxidants & Redox Signaling* 11:2, 241-249. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 20. Po Sing Leung, Yuk Cheung Chan. 2009. Role of Oxidative Stress in Pancreatic Inflammation. *Antioxidants & Redox Signaling* 11:1, 135-166. [Abstract] [Full Text PDF] [Full Text PDF with Links]

- 21. R.X. Santos, S. Cardoso, S. Silva, S. Correia, C. Carvalho, J. Crisóstomo, L. Rodrigues, C. Amaral, T. Louro, P. Matafome, M.S. Santos, T. Proença, A.I. Duarte, R. Seiça, P.I. Moreira. 2009. Food Deprivation Promotes Oxidative Imbalance in Rat Brain. *Journal of Food Science* 74:1, H8-H14. [CrossRef]
- 22. Stephen J Duffy, Anthony M Dart. 2008. Novel cardiac therapies and innocent bystanders. The Lancet 371:9626, 1726-1728. [CrossRef]
- 23. Muhammad Iqbal, Syed Shah, Sonalis Fernandez, Jocelyne Karam, Girardin Jean-Louis, Samy I. McFarlane. 2008. Obesity, obstructive sleep apnea, and cardiovascular risk. *Current Cardiovascular Risk Reports* 2:2, 101-106. [CrossRef]
- 24. Raymond M Schiffelers, Marcel Ham Fens, Janneke M van Blijswijk, Dieuwertje I Bink, Gert Storm. 2007. Targeting the retinal microcirculation to treat diabetic sight problems. *Expert Opinion on Therapeutic Targets* 11:11, 1493-1502. [CrossRef]
- 25. Melvin R. Hayden, James R. Sowers. 2007. Redox Imbalance in Diabetes. *Antioxidants & Redox Signaling* **9**:7, 865-867. [Citation] [Full Text PDF] [Full Text PDF with Links]